# Cytokine TRAIL death receptor agonists: design strategies and clinical prospects

Anne V. Yagolovich,<sup>a</sup>\* <sup>(0)</sup> Marine E. Gasparian,<sup>b</sup> <sup>(0)</sup> Alina A. Isakova,<sup>a,b</sup> <sup>(0)</sup> Artem A. Artykov,<sup>b</sup> <sup>(0)</sup> Dmitry A. Dolgikh,<sup>a,b</sup> <sup>(0)</sup> Mikhail P. Kirpichnikov<sup>a,b</sup> <sup>(0)</sup>

<sup>a</sup> Faculty of Biology, Lomonosov Moscow State University, 119234 Moscow, Russia

<sup>b</sup> Shemyakin–Ovchinnikov Institute of Bioorganic Chemistry of the Russian Academy of Sciences,

117997 Moscow, Russia

The development of therapeutic bispecific antibodies and hybrid proteins is one of the most urgent biomedical technologies with obvious clinical prospects. At the same time, advanced strategies of molecular design of drugs with new properties are coming to the forefront. The tumour necrosis factor-related apoptosis inducing ligand (TRAIL) receptor pathways are important components of the immune system involved in the immune surveillance and selective elimination of transformed cells. TRAIL-based proteins are therefore promising drug candidates for the treatment of malignant tumours and autoimmune diseases. In the first series of clinical trials, drugs targeting the death receptors DR4 or DR5, did not show significant anti-cancer activity. This is due to the TRAIL resistance mechanisms that tumours evolve to evade the efficient induction of apoptotic signalling. However, a wide range of novel TRAIL death receptor-targeted formulations are currently being developed, mainly to improve stability, enhance death receptor clustering and involve additional tumour targets. Over the past decade, several dozens of multitargeted fusion proteins with either TRAIL protein or DR5-specific agonistic monoclonal antibodies have been developed to improve therapeutic efficacy. These include fusions with either short peptide tags or large functional proteins, as well as antibody fragments targeting molecular pathways involved



in angiogenesis or proliferative signalling such as epidermal growth factor receptor (EGFR), vascular endothelial growth factor receptor (VEGFR), programmed death-ligand 1 (PD-L1), *etc.* Collectively, these multimodal proteins enhance the activation of extrinsic and intrinsic apoptotic pathways in transformed cells, as well as affect the tumour microenvironment. This comprehensive review systematizes the bispecific and multivalent fusion proteins and conjugates targeting TRAIL death receptors, analyze the molecular mechanisms by which they overcome tumour resistance to TRAIL, and assess their clinical prospects. The bibliography includes 236 references.

1

2

7

9

Keywords: TRAIL, fusion proteins, death receptors DR4 and DR5, cancer therapy.

### Contents

- 1. Introduction
- 2. TRAIL signalling pathways
- 3. Overcoming cancer cell resistance to TRAIL-induced apoptosis 3
- 4. TRAIL fusions and conjugates aimed at increasing antitumour 5 efficiency by improving protein stability and pharmacokinetic parameters
- 5. Fusions aimed at increased clustering of the death receptors DR4 and DR5
- 6. Bifunctional fusions of TRAIL with antitumour peptides or proteins

### 1. Introduction

Due to their ability to simultaneously activate several signalling pathways that affect tumour development, bispecific fusion proteins are on the cutting-edge of the modern anticancer drug discovery.<sup>1</sup> Among them, cytokines engineered for the selective cell targeting, including so-called 'supercytokines', 'immunocytokines', 'fusokines' and other synthetic cytokines, serve as an important platform for the development of novel

7. TRAIL fusions with antibody fragments targeting	11
8. Multimeric and bifunctional antibodies targeting TRAIL	16
9. Clinical trials of next generation TRAIL death receptor	19
agonists 10. Conclusion	20
11. List of abbreviations	20
12. References	21

protein-based the rapeutics with enhanced biological properties.  $^{2}\$ 

The cytokine TRAIL, a member of the tumour necrosis factor (TNF) family, is an important component of the immune system that selectively eliminates transformed and aberrant cells by apoptosis after binding to the death receptors (DRs) DR4 or DR5.<sup>3,4</sup> Unlike TNF and FAS ligands, TRAIL is the only natural cytokine that induces apoptosis in transformed cells without significant toxicity to normal tissues.<sup>3,5</sup> This has served as the

basis for the development of DR agonists for the targeted therapy of tumour diseases. However, the first-generation DR agonist, soluble recombinant TRAIL alone or in combination with chemotherapeutic drugs, appeared to be ineffective in the clinic.<sup>5–11</sup> Similarly, the first monoclonal antibodies targeting DRs have also shown very limited anti-cancer effect.<sup>12</sup>

Like some other members of the TNF family, TRAIL can induce the production of pro-inflammatory chemokines and cytokines that promote a tumour-supportive immune microenvironment, which may counteract its antitumour activity.13 There is also increasing evidence that the TRAIL DR signalling pathway is involved in the regulation of cancer invasion and metastasis, with both positive and negative roles being reported.<sup>14–16</sup> This is achieved by a complex balance between TRAIL-induced activation of pro-apoptotic signal transduction and non-canonical pro-survival pathways.<sup>17,18</sup> However, despite the failure of the first-generation DR agonists, the implementation of rational and sophisticated molecular design strategies has led to a new wave of promising developments, comprising of several dozen new molecular constructs targeting TRAIL DRs with enhanced agonistic improved pharmacokinetics and therapeutic activity, efficacy.<sup>17,19–24</sup> This review aims to provide a comprehensive overview of the developed bispecific and multivalent fusion proteins and conjugates targeting TRAIL DRs with an assessment of their clinical prospects.

#### 2. TRAIL signalling pathways

The cytokine TRAIL was identified by two independent groups in 1995–1996 on the basis of its sequence homology to TNF and FasL.<sup>25,26</sup> Analysis of the crystal structure of the TRAIL protein revealed that soluble TRAIL forms a homotrimer similar to other members of the TNF family.<sup>27</sup> The biological activity of TRAIL is critically dependent on the presence of an unpaired cysteine (Cys230), which is involved either in the formation of an interchain disulfide bridge, resulting in the formation of a poorly active TRAIL dimer, or in the formation of an active trimer by chelation of a single zinc atom.<sup>28,29</sup> TRAIL is expressed as a type II transmembrane protein, and its extracellular domain can be proteolytically cleaved from the cell surface presumably by a cysteine proteases to form a soluble ligand.<sup>30,31</sup> As soluble TRAIL has shown high biological activity, the bulk

**A.V.Yagolovich**. Candidate of Biological Sciences, Assistant at the Department of Bioengineering, Faculty of Biology, Lomonosov Moscow State University.

E-mail: yagolovichav@my.msu.ru

*Current research interests*: molecular mechanisms of cell death, mechanisms of tumour resistance, antitumour therapy, antitumour immune response, protein engineering, structure and functions of proteins, nanoscale delivery systems.

**M.E.Gasparian**. Candidate of Biological Sciences, Senior Researcher, Laboratory of Protein Engineering, Shemyakin– Ovchinnikov Institute of Bioorganic Chemistry RAS.

E-mail: marine\_gasparian@yahoo.com

*Current research interests*: protein engineering, structure and functions of proteins, molecular mechanisms of cell death, development of protein drugs for biomedical purposes.

A.A.Isakova. Junior Researcher, Laboratory of Protein Engineering, Shemyakin–Ovchinnikov Institute of Bioorganic Chemistry RAS. E-mail: alina.labbio@gmail.com

*Current research interests*: protein engineering, structure and functions of proteins, development of protein drugs for biomedical purposes.

of the studies have been carried out using recombinant preparations of the extracellular domain of the protein containing 95-281 or 114-281 amino acids.

To date, four membrane-bound receptors have been identified, TRAIL-R1 (DR4), TRAIL-R2 (DR5), TRAIL-R3 (DcR1), and TRAIL-R4 (DcR2), which bind TRAIL with similar affinities and subnanomolar dissociation constants.<sup>29,32,33</sup> The agonistic death receptors (DRs) DR4 and DR5 are type I transmembrane proteins and contain an intracellular death domain (DD) that normally stimulates apoptosis upon TRAIL binding.34,35 DcR1 and DcR2 are transmembrane proteins that do not have a fully developed intracellular DD and are unable to transmit an apoptotic signal, so they are called decoy receptors.<sup>36-39</sup> In addition, decoy receptors compete with the apoptosis-inducing DR4 and DR5 for ligand binding and inhibit TRAIL-induced apoptosis.<sup>40</sup> TRAIL also showed high affinity for the secreted soluble decoy receptor osteoprotegerin (OPG), which lacks both transmembrane and cytoplasmic residues and attenuates TRAIL-induced apoptosis.41 The guide tree and schematic structure of human TRAIL receptors are shown in Figure 1.

Binding of TRAIL homotrimer induces trimerization of DR4 and DR5, leading to the assembly of the death-inducing signalling complex (DISC) and subsequent recruitment of the adaptor protein Fas-associated death domain (FADD).<sup>42</sup> The latter acts as a bridge between the death receptor complex and the caspase-8 initiator prodomain. Upon activation of the extrinsic cell death pathway, dimerization of caspase-8 in the DISC leads to the formation of mature caspase-8, which activates immediate downstream effector caspases such as caspase-3, -6, and -7, triggering apoptosis.<sup>43</sup> The catalytically inactive caspase-8-homologous protein c-FLIP (FLICE-like inhibitory protein) can compete with caspase-8 for binding to FADD preventing active DISC formation and mediating cell resistance to apoptosis.<sup>44,45</sup>

TRAIL binding to DRs is critical for activation of the extrinsic apoptotic pathway, and downregulation of DR4 and DR5 is sufficient to render cancer cells resistant to TRAIL.<sup>46</sup> It has previously been shown that DR4 and DR5 can undergo spontaneous and ligand-mediated endocytosis and recycling independent of cancer cell sensitivity to TRAIL.<sup>47</sup> Disruption of receptor trafficking by dysfunctional cargo transporter proteins and nuclear translocation signalling proteins can inhibit

**A.A.Artykov**. Junior Researcher, Laboratory of Protein Engineering, Shemyakin–Ovchinnikov Institute of Bioorganic Chemistry RAS. E-mail: art.al.artykov@gmail.com

*Current research interests*: protein engineering, instrumental methods for obtaining molecular biological data.

**D.A.Dolgikh**. Doctor of Biological Sciences, Professor, Head of the Laboratory of Protein Engineering, Shemyakin–Ovchinnikov Institute of Bioorganic Chemistry RAS, Professor of the Department of Bioengineering, Faculty of Biology, Lomonosov Moscow State University. E-mail: dolgikh@nmr.ru

*Current research interests*: protein engineering, structure and functions of proteins, development of biotechnological protein drugs for biomedical purposes.

M.P.Kirpichnikov. Doctor of Biological Sciences, Professor, Academician of Russian Academy of Sciences, Dean of the Faculty of Biology, Lomonosov Moscow State University, Head of the Department of Bioengineering, Faculty of Biology, Lomonosov Moscow State University, Head of Department of Bioengineering, Shemyakin–Ovchinnikov Institute of Bioorganic Chemistry RAS. E-mail: kirpichnikov@inbox.ru

*Current research interests*: physical and chemical biology, protein engineering, biotechnology.



Figure 1. The guide tree and schematic structure of human TRAIL receptors. Sequences were aligned using Uniprot with the Clustal Omega program following Guide tree option.

translocation of DR from the trans-Golgi network (TGN) to the plasma membrane, resulting in reduced surface expression of DR.<sup>46</sup> In addition to localization on the plasma membrane, DR4 and DR5 are found in the cytoplasm and nucleus.<sup>48</sup> It has been shown that TRAIL can induce nuclear translocation and chromatin localization of DR4 and DR5 highlighting an additional role for surface-activated DRs *via* direct trafficking and signalling inside the nucleus.<sup>49</sup> Two functional nuclear localization signals have been identified in the DR5 receptor sequence, which, when mutated, together with knockdown of importin 1 by siRNA, prevented the nuclear localization of DR5, leading to increased DR5 expression on the cell surface and sensitization of HeLa and HepG2 cells to TRAIL.<sup>50</sup> Recent studies have identified specific tumour-promoting functions of nuclear DR5 as a regulator of let-7 maturation.<sup>51</sup>

In so-called type II cells, TRAIL can trigger the intrinsic apoptotic pathway, where activated caspase-8 cleaves the proapoptotic BH3-only BCL-2 family protein Bid. Truncated Bid (tBid), in turn, activates the proapoptotic proteins Bak and Bax, leading to mitochondrial outer membrane permeabilization (MOMP) and cytochrome C release.<sup>52</sup> Subsequently, the apoptotic protease activating factor 1 (APAF1) protein binds to cytochrome C and activates caspase-9. At the same time, during MOMP, a SMAC/DIABLO (second mitochondrial caspase activator) is released, which blocks the activity of inhibitors of apoptosis proteins (IAPs), thereby promoting apoptosis.<sup>53</sup> Active caspase-9 activates caspase-3, -6 and -7 in the same way as caspase-8. Anti-apoptotic members of the BCL-2 protein family, including BCL-2, BCL-xL, BCL-w, and MCL1, negatively regulate the TRAIL-mediated intrinsic death pathway by preventing MOMP.54

In addition to apoptosis, TRAIL death receptors can induce non-canonical signalling, leading to tumour resistance.<sup>18,55</sup> Activation of pro-inflammatory, pro-survival and proliferation pathways leading to pro-tumourigenic and, in some cases, metastasis-promoting effects following TRAIL treatment *in vitro* and *in vivo* have been described.<sup>17,56–58</sup> Non-canonical signalling is mediated by the formation of a secondary signalling complex that includes, in addition to FADD and caspase-8, RIPK1 (serine/threonine protein kinase 1), TRAF (TNF receptor-associated factor 2) and TRADD (TNF receptor-associated death domain).<sup>56</sup> This signalling complex can then activate various pro-tumourigenic pathways, including PI3K, MAPK/ERK, I $\kappa$ B/NF- $\kappa$ B, STAT3, Akt, Src and JAK2.

Thus, activation of non-canonical TRAIL pathways is a challenge not only because of the development of tumour resistance to DR agonists, but more importantly of the potential for tumour dissemination and metastasis.

A graphical representation of the TRAIL signalling pathways is shown in Figure 2.

# **3.** Overcoming cancer cell resistance to TRAIL-induced apoptosis

Despite the high antitumour activity of TRAIL, it can also induce acquired resistance of tumour cells to apoptosis, which is considered an obstacle to its clinical use.<sup>59</sup> As a result, early clinical trials with several variants of TRAIL death receptor agonists have shown only limited therapeutic effect. Some of them showed insufficient antitumour activity, while for others, signs of toxicity were observed. Combination with various chemotherapeutic agents has not significantly improved the therapeutic potential of DR agonists, either because of a lack of appropriate combination therapy, or because of increased acquired cell resistance in response to the combination treatment.<sup>21</sup> It should be noted that many cancer cells are inherently resistant to TRAIL due to dysregulation of antiapoptotic proteins such as BCL-2, BCL-XL, Bfl1/A1, c-FLIP, cIAP, survivin, Mcl-1 and XIAP.18 To overcome TRAIL resistance, various chemotherapeutic agents and natural compounds have been proposed for combination therapy.<sup>60</sup> A number of synthetic small molecules have been obtained that suppress the activity of anti-apoptotic proteins and re-sensitize cancer cells to TRAIL.<sup>22</sup> To date, the most promising agents for combination therapy to overcome TRAIL resistance are proteasome inhibitors, BH3-mimetics, and cyclin-dependent



**Figure 2.** TRAIL signalling pathways. (*a*) Apoptosis pathway. Activation of DR4 and DR5 by TRAIL induces the extrinsic apoptosis pathway (left). The intrinsic pathway (right) is activated by a variety of stimuli and leads to the release of proapoptotic proteins from the mitochondria. The two pathways interact *via* caspase-8, which, once activated by the DRs, can cleave BID, further activating the intrinsic pathway, and conversely, caspase-3 can cleave and activate caspase-8 in a feedback loop, thus amplifying the apoptotic signal. (*b*) TRAIL-mediated non-apoptotic pathways. Anti- or pro-survival mechanisms appear to be context-dependent. DISC is death-inducing signaling complex; cFLIP is cellular FLICE [FADD-like IL-1 $\beta$ -converting enzyme]-inhibitory protein; SMAC is second mitochondrial activator of caspases; Cyt C is cytochrome C; XIAP is X-linked inhibitor of apoptosis. TRAF2 is TNF receptor-associated factor 2; NEMO is NF-kappa-B essential modulator; NF- $\kappa$ B is nuclear factor- $\kappa$ B; RIPK1 is receptor-interacting serine/threonine-protein kinase 1; JNK is C-Jun N-terminal kinase; ERK is extracellular signal-regulated kinase, BAD is BCL-2-associated agonist of cell death; BIM is BCL-2-interacting mediator of cell death; PUMA is p53-upregulated modulator of apoptosis; NOXA is phorbol-12-myristate-13-acetate-induced protein 1; p38-BID is BH3-interacting domain death agonist; BAK is BCL-2/Killer 1 antagonist; BAX is BCL-2-associated X protein.

kinase 9 (CDK9) inhibitors. Bortezomib is a selective 26S proteasome inhibitor approved by the US Food and Drug Administration (FDA) in 2003 under the trade name Velcade. It has demonstrated synergistic effects in various cancer cells when combined with DR agonists by increasing DR5 receptor expression and enhancing DISC formation.<sup>61–63</sup> Despite promising preclinical results, the combination of first-generation DR4 and DR5 agonistic antibodies, mapatumumab and conatumumab, correspondingly, with bortezomib failed to show additional therapeutic benefit (clinical trials NCT00315757, NCT00791011).

The BH3-only proteins initiate apoptosis by neutralizing the pro-survival BCL-2 family of proteins. Over the past 20 years, various small molecules called BH3-mimetics have been synthesized that mimic the function of BH3-only proteins and destroy cancer cells.<sup>64</sup> BH3-mimetics and TRAIL are highly synergistic in inducing apoptosis.<sup>65,66</sup> For example, a novel multimeric anti-DR5 IgM agonist antibody IGM-8444 (discussed further in Section 8) synergizes *in vitro* and *in vivo* with the BCL-2 selective inhibitor ABT-199 (also known as venetoclax, the only BH3 mimetic that has been approved by the FDA).<sup>67</sup>

Cyclin-dependent kinase 9 is a potential therapeutic target in cancer, as its overexpression correlates with cancer progression and poor clinical outcomes. CDK9 regulates multiple cellular functions including proliferation, survival, cell cycle regulation, DNA damage repair and metastasis. For this reason, many specific small molecule inhibitors have been developed over the past decades.<sup>68</sup> The novel, orally bioavailable CDK9 inhibitor Atuveciclib induced significant apoptosis in pancreatic cancer cell lines in combination with TRAIL *via* concomitant suppression of cFLIP and Mcl-1.<sup>69</sup> The clinically advanced CDK9-targeting drug Dinaciclib significantly enhanced the anticancer properties of TRAIL, regardless of the sensitivity or resistance of cancer cells to chemotherapy or targeted therapy.<sup>70,71</sup>

Dordaviprone (ONC201) is a first-in-class small molecule dopamine D2 receptor (DRD2) antagonist that upregulates TRAIL and death receptor 5 (DR5) expression by inactivating the pro-survival Akt/ERK kinases, thereby demonstrating potent antitumour activity in multiple cancer types.<sup>72</sup>

Recently, ONC201 monotherapy was shown to be well tolerated and to have long-term and clinically meaningful efficacy in recurrent diffuse midline glioma with H3 K27M mutation.<sup>73</sup> Other ONC201 analogues, designated ONC206 and ONC212, are under intensive *in vivo* and *in vitro* investigation, and have shown comparable or superior antitumour activity to ONC201.<sup>74,75</sup> The combination of imipridones ONC201 or ONC206 with temozolomide and radiotherapy also reduced intracranial tumour burden and prolonged survival in an orthotopic mouse model of IDH (isocitrate dehydrogenase) wild-type glioblastoma.<sup>76</sup>

Resistance of cancer cells to DR agonists may also be due to overexpression of members of the inhibitor of apoptosis protein (IAP) family. Drugs called SMAC mimetics have been developed that mimic natural IAP antagonists such as the second mitochondrial activator of caspase (SMAC).<sup>77</sup> The combination of such molecules with TRAIL overcomes cancer cell resistance

either by degrading cIAP1 or XIAP, or by suppressing cFLIP(L).  $^{78-80}$ 

#### 4. TRAIL fusions and conjugates aimed at increasing antitumour efficiency by improving protein stability and pharmacokinetic parameters

To overcome the aforementioned obstacles, several modification strategies have been developed to improve the antitumour properties of TRAIL death receptor agonists (Fig. 3).

Increasing the serum stability of TRAIL and prolonging its circulation in blood is a viable strategy to improve its antitumour efficacy. It is well known that the serum half-life of TRAIL is as low as 1 h.<sup>8</sup> Due to fast elimination from the body *via* renal clearance and poor pharmacokinetic profile, TRAIL signalling is insufficient to induce apoptosis in cancer cells. Unsatisfactory stability and pharmacodynamics have been among the challenges, which hampered the clinical translation of TRAIL-based therapies.<sup>81</sup> Recombinant soluble TRAIL can exist in different oligomeric states.<sup>82,83</sup> The hyper-oligomerized forms of TRAIL demonstrated extremely high potency in inducing apoptosis. Therefore, the oligomeric state of TRAIL can influence its stability and biological activity.<sup>84</sup>

One of the first promising TRAIL modifications was leucine or isoleucine zipper-TRAIL.<sup>85–89</sup> The unique structural modification of zipper-TRAIL involves the introduction of hydrophobic interactions and intermolecular forces that promote self-assembly into stable multimeric structures.<sup>88</sup> These structures often adopt a 'zipper-like' pattern, where the TRAIL molecules align and bind together in a highly organized manner. Studies in breast, prostate and lung carcinomas have shown that self-assembly not only increases the stability of Zipper-TRAIL but also contributes to its increased resistance to proteolytic degradation. These stable structures not only protect Zipper-TRAIL from rapid degradation in the bloodstream, but also have



Figure 3. Schematic representation of fusion proteins targeting TRAIL death receptor pathway. Fusions of TRAIL with moieties for trimer stabilization, antitumour peptides, antibody fragments, or death receptor agonistic bispecific antibodies were designed to improve stability, pharmacokinetics, and antitumour efficacy.

thereby promoting the apoptotic signalling. A number of chemical modifications has been used to improve the therapeutic capacity of TRAIL. One such modification is the chemical attachment of polyethylene glycol (PEG). PEG is a non-immunogenic biological compound consisting of repeating units of ethylene glycol. Covalent and non-covalent attachment of PEG to proteins increases half-life, reduces immunogenicity and improves solubility and stability of therapeutic drugs.<sup>90</sup> According to this strategy, an N-terminally PEGylated TRAIL derivative (PEG-HZ-TRAIL) with trimerforming zipper sequences has been constructed.91 Site-specific PEGylation of the  $\mathrm{NH}_2$  terminus of HZ-TRAIL with methoxypoly(ethylene glycol) aldehyde (mPEG-ALD) was carried out by reductive amination in the presence of sodium cyanoborohydride at acidic pH. Due to the relatively lower pKa values of the NH<sub>2</sub>-terminal amine (a-amino group) compared to that of the internal lysine residues (of  $\varepsilon$ -amino groups), the reductive amination is quite selective for N-terminal-specific PEGylation. In vivo experiments in a colorectal carcinoma model showed increased stability of PEG-HZ-TRAIL due to its protection from proteolytic degradation in the bloodstream. PEGylation masks the TRAIL surface, reducing its vulnerability to enzymatic cleavage. In additions, the prolonged circulation and improved pharmacokinetics result in increased DR binding on cancer cells, leading to more efficient apoptotic signalling. Further, this PEGylated long-acting recombinant human TRAIL has been shown to ameliorate liver fibrosis and cirrhosis by selectively depleting activated hepatic stellate cells<sup>92</sup> and to induce DR-mediated apoptosis in collagen-producing myofibroblasts through upregulated DR5, reversing established skin fibrosis in scleroderma.93

In another work, site-specific PEGylation was performed using click chemistry approach by covalently binding PEGmaleimide to the TRAIL/N109C mutant bearing N-terminal cysteine residue. mPEG<sub>MAL</sub>-N109C showed improved in vitro stability and greater therapeutic potential in a tumour xenograft model, with better drug delivery and bioavailability compared to the TRAIL N-terminally PEGylated using mPEG-ALD.94 The TRAIL/N109C variant was used to produce N109C-vcMMAE and PEG-TRAIL-vcMMAE conjugates with a cathepsincleavable linker between TRAIL and monomethyl auristatin E (MMAE).<sup>95,96</sup> MMAE is a potent antimitotic agent that inhibits cell division by blocking the polymerization of tubulin. Upon receptor binding, these fusions rapidly internalize into the cytoplasm of cancer cells and release MMAE into the lysosome via lysosomal-specific linker cleavage to induce growth arrest and cell death of cancer cells via apoptosis. Both fusions showed high antitumour activity and improved pharmacokinetic parameters. The half-life of PEG-N109C-vcMMAE in the rat was prolonged to 11.54 h compared to TRAIL/N109C (1.91 h) and N109C-vcMMAE (4.26 h).

One promising approach is to use trimerization domains derived from various proteins to enhance the stability of target proteins. These domains can be employed to promote the self-association of monomeric proteins into trimeric structures, which is often critical for the expression of their biological functions. The trimerization domain (TD) of human collagen, characterized by a repeating Gly-X-Y motif, imparts stability to proteins through its ability to self-assemble into a triple-helical structure that mimicks the structure of native collagen. TRAIL-TD fusion halved the IC<sub>50</sub> in small-cell lung cancer cell model.<sup>97</sup> Another way to stabilize TRAIL is to fuse it to the Trimer-Tag, a human C-propeptide of alpha1(I) collagen. A recombinant

fusion protein, SCB-313, consisted of Trimer-tag fused to the TRAIL C-terminus, allowing TRAIL to form a stable covalently linked homotrimer. SCB-313 had an extended half-life in the circulation due to its resistance to enzymatic cleavage and rapid clearance. The improved pharmacokinetic profile ensured sustained bioavailability, allowing for a longer therapeutic window and enhanced antitumour activity in colon carcinoma models *in vitro* and *in vivo*. The trimeric configuration not only stabilized the protein, but also increased its binding affinity to death receptors on cancer cells, resulting in more potent apoptotic signalling and increased cancer cell death.<sup>98</sup>

Another stabilizing modification of soluble TRAIL is fusion with an adenovirus knobless fibre motif. Three engineered TRAIL variants, designated FA1FT, HA5FT and HA5ST, were created by fusing the avian Ad1 spineless fibre (FA1FT), the N-terminal tail and the first two repeats of the human Ad5 fibre shaft (HA5FT) and the last repeat of the human Ad5 fibre shaft (HA5ST) to the N-terminus of TRAIL. These constructs were designed to form a trimeric configuration in which the three TRAIL monomers are linked to each other *via* the adenovirus fibre rod domain. Among these proteins, HA5ST showed the highest anticancer activity and improved pharmacodynamics.<sup>99</sup>

It has also been shown that improved TRAIL stability can be achieved by genetically fusing TRAIL to the albumin binding domain (ABD).<sup>100</sup> The ABD component of the ABD-hTRAIL fusion promoted binding to albumin in the bloodstream, which significantly prolonged the half-life, making ABD-hTRAIL more stable than native TRAIL. The presence of ABD protected TRAIL from proteolytic degradation, reducing its susceptibility to enzymatic cleavage and denaturation. This protein has been tested in vivo in models of colon carcinoma and has shown a remarkable ability to induce cancer cell death, resulting in tumour growth inhibition. The same group of researchers created a fusion in which the IgG-binding affibody, IgBD, was genetically fused to the N-terminus of TRAIL to form IgBD-TRAIL, and compared its cytotoxicity, serum half-life, and antitumour activity with ABD-TRAIL.101 The serum halflife of both fusions exceeded that of TRAIL but the antitumour effects of the intravenously injected IgBD-TRAIL were superior to that of ABD-TRAIL.

Aiming to improve the TRAIL stability, another research group<sup>102</sup> has fused the crystallizable region of a human IgG1 fragment (Fc) with the TRAIL N-terminus to produce an Fc-TRAIL fusion. The chimeric protein had a significantly longer half-life in mice and was more effective than TRAIL in inhibiting tumour growth in a xenograft model of lung cancer.

A strategy combining increased stability and additional antitumour activity was also implemented by fusing TRAIL to a hexameric arginine deiminase (ADI). ADI-TRAIL benefited from the structural and functional synergy between the two moieties and had an extended half-life *in vivo*. It not only activated the DR5 receptor, but also suppressed survivin, and sensitized cancer cells to TRAIL-induced apoptosis significantly inhibiting tumour growth in colorectal carcinoma models.<sup>103</sup>

An interesting modification of recombinant mutant human TRAIL (rmh TRAIL) developed by Beijing Sunbio Biotech and further renamed CPT (circularly permuted TRAIL, drug name Aponermin) consists of the N-terminal TRAIL amino acids (121-135) fused with a flexible linker Gly<sub>5</sub> to the C-terminal TRAIL amino acids (135-281).<sup>104</sup> CPT has potent antitumour activity *in vitro* and *in vivo*, and has demonstrated the clinical activity in a Phase II trial in patients with relapsed or refractory multiple myeloma as a single agent<sup>105</sup> and in a Phase III trial in combination with thalidomide and dexamethasone.<sup>106</sup>

To optimize the oligomerization, a Flag tag and chicken tenascin-C (TNC) oligomerization domain (110-139 amino acids) were fused to the N-terminus of TRAIL to obtain Flag-TNC-TRAIL.<sup>107</sup> Introduction of the TNC domain enhanced TRAIL activity after secondary cross-linking: oligomerized Flag-TNC-TRAIL strongly induced cell death in myeloma cells, but also activated pro-inflammatory signalling pathways.<sup>108</sup> An alternative way to force trimerization is a covalently linked TRAIL trimer (TR3) created by genetically fusing three consecutive extracellular domains of TRAIL in a head-to-tail configuration. This molecular design provided improved stability without altering the native killing capacity of TRAIL. The authors also generated an scFv-TR3 fusion of a single-chain antibody fragment (scFv) specific for murine red blood cells (RBCs) with the NH2-terminus of TR3. Similarly, scFv-S-TR3 variant with an elongated spacer domain between the targeting scFv and TR3 comprising four globular domains of the human complement regulatory proteins decay accelerating factor (DAF, CD55) and complement receptor 1 (CR1, CD35) was produced to anticipate the possible steric constraints. RBCs coated with the scFv-S-TR3 have demonstrated high potential for clinical cancer therapy in pancreatic adenocarcinoma models.<sup>109</sup>

A TRAIL-Mu3 fusion protein was obtained to enhance the membrane permeability of TRAIL by replacing the amino acid residues VRERGPQR (114–121) with RRRRRRR.<sup>110,111</sup> TRAIL-Mu3 more effectively activated the caspase cascade in pancreatic cancer cell lines and demonstrated increased efficacy in preclinical models of pancreatic cancer compared to wild-type TRAIL (wtTRAIL).

Another original approach was used to generate biologically functional DR4 and DR5 agonists (KD413, KD 506 and KD548) from a Kringle domain (KD) scaffold by generating a synthetic KD library on the surface of yeast cells and randomizing 45 residues in the loops of the human KD template.<sup>112</sup> The KD variants selected against DR4 and DR5 showed antitumour activities *in vitro* and *in vivo*. In addition, using a loop grafting technique, the authors created a bispecific kringle domain agonist bvKD548-55 as a potent agonist of death receptors 4 and 5.<sup>113</sup>

# 5. Fusions aimed at increased clustering of the death receptors DR4 and DR5

One of the reasons for the limited efficacy exhibited by recombinant soluble trimeric TRAIL is its insufficient receptor

clustering capacity during short-term presence in the bloodstream. Since the efficacy of DR agonists in inducing tumour cell apoptosis is highly dependent on their valency, therefore, recent efforts have focused on improving DR clustering to enhance antitumour activity. A potent TRAIL receptor agonist APG350 with a hexavalent binding pattern that allows clustering of six TRAIL receptors per drug molecule, was produced by C-terminal fusion of an engineered IgG1-Fc to a single-chain TRAIL receptor binding domain (scTRAIL-RBD).<sup>114</sup> Antitumour efficacy of APG350 exceeded that of soluble TRAIL in pancreatic cancer with limited recurrent tumour growth and metastases.<sup>115</sup> Subsequently, an APG350 fusion derivative ABBV-621 was created containing a single IgG1-Fc point mutation (asparagine 297 to serine) that effectively removes the glycosylation site to eliminate binding to all Fcy receptors and complement component C1q. Preclinical studies have demonstrated improved pharmacokinetics and antitumour activity of ABBV-621, particularly when combined with chemotherapeutic agents such as docetaxel, irinotecan, and the selective BCL-XL inhibitor A-1331852, in preclinical models of colorectal and pancreatic cancer.<sup>116</sup>

Novel genetic engineering techniques using small molecule superglues to covalently ligate proteins offer promising alternatives for protein oligomerization. Using the molecular superglue SpyTag/SpyCatcher to create more complex TRAIL variants, such as hexameric TRAIL (HexaTR), resulted in a significant increase in apoptosis induction.<sup>117</sup> The development of albumin-binding HexaTR (ABD-HexaTR) with an extended serum half-life demonstrated its remarkable antitumour activity in vivo, effectively eliminating various xenograft tumours. These data suggest that the superglue-mediated higher-order assembly approach is valuable for improving proapoptotic TRAIL signalling and has significant potential for advancing DR agonists in cancer therapy. The same research group<sup>118</sup> further generated hexavalent fusion SnHexaTR by catalyzing trivalent TRAIL variants with N-terminal fusion of SnoopTagJr/SnoopDogTag with Snoopligase. The in vitro cytotoxicity of SnHexaTR was 10-40 times greater than that of TRAIL in several tumour cell lines. SnHexaTR showed a longer serum half-life and greater tumour uptake than TRAIL, resulting in enhanced cytotoxicity and antitumour activity of TRAIL.

A summary of TRAIL fusions aimed at improving protein stability and pharmacokinetic parameters is presented in Table 1.

Table 1. TRAIL fusions and conjugates to improve protein stability and pharmacokinetic parameters.

Name	Structure	Effects	In vivo model	Ref.
LZ-huTRAIL LZ-muTRAIL	Leucine zipper (LZ) fused with human or murine TRAIL	Improved pharmacodynamics and antitumour effect	Xenograft colon carci- nomas in SCID mice	85
LZ-TRAIL	Modified yeast GCN4-pII LZ motif fused to N-terminus of TRAIL (120–281 aa)	Improved pharmacodynamics and antitumour effect	Xenograft breast carcinoma in mice	87
ATF-TRAIL	Human leucine zipper peptide ATF7-pII fused to the N-terminus of TRAIL	Increased stability and binding to death receptors, improved antitumour apoptotic activity	Orthotopic breast cancer xenograft model	88
izTRAIL	Isoleucine zip motif (IEKKIEA) <sub>4</sub> fused with TRAIL	Considerably increased activity in confluent cultures	-	89
PEG-HZ-TRAIL	Site-specific NH <sub>2</sub> -terminal PEGylation of a TRAIL variant possessing trimer-forming zipper sequences	Increased stability, pharmacokinetics and antitumour efficacy	Colorectal cancer xenograft model	91
mPEG <sub>MAL</sub> -N109C	Covalently PEGylated TRAIL N109C mutant by PEG-maleimide	Higher therapeutic potential compared to N-terminally PEGylated TRAIL	Lung carcinoma xenograft model	94

Nomo	Structure	Effects	Lu vivo modol	Def
Name	Structure	Effects	In vivo model	Kei.
N109C-vcMMAE PEG-TRAIL- vcMMAE	TRAILN109C variant conjugated with MMAE or PEG and MMAE	Improved pharmacokinetics and potent antitumour activity	Lung carcinoma and pancreatic cancer xenograft model	95, 96
TRAIL-TD	Human collagen XVIII trimerization domain (TD) fused to C-terminus of TRAIL	High apoptotic activity	-	97
SCB-313	Human C-propeptide of α1(I) collagen (Trimer-Tag) at the C-terminus of human TRAIL	Enhanced pharmacokinetics and antitumour efficacy	Colorectal cancer xenograft model	98
HA5ST	C-terminal shaft repeat of the human adenovirus type 5 (Ad5) fiber protein fused with TRAIL	Improved pharmacokinetics, bio- availability and antitumour activity	Breast cancer xenograft model	99
ABD-hTRAIL	TRAIL with an albumin-binding domain (ABD) at the N-terminus	Prolonged circulation, improved binding affinity, enhanced antitumour effect	Colorectal cancer xenograft model	100
IgBD-TRAIL	An IgG-binding affibody, IgBD genetically fused to the N-terminus of TRAIL	Prolonged serum half-life, significantly enhanced antitumour effect	Colorectal cancer xenograft model in mice	101
Fc-TRAIL	The Fc portion of human IgG1 fused to the N-terminus of human TRAIL	Prolonged half-life in mice and more effective in inhibiting tumour growth compared with TRAIL	Lung cancer xenograft model	102
ADI-TRAIL	Arginine deiminase (ADI) fused to N-terminus of TRAIL	Extended half-life, sensitization of cancer cells to TRAIL, enhanced antitumour activity	Colorectal cancer xenograft model	103
CPT (circularly permuted TRAIL)	The N-terminal amino acids 121–135 of TRAIL connected to the C-terminal amino acids 135–281 by a flexible linker	Improved pharmacodynamics and antitumour effect	Lung cancer xenograft model in mice	104
Flag-TNC-hTRAIL	Flag tag and tenascin-C (TNC) oligomerization domain fused with TRAIL	Enforced covalent trimerization and increased apoptotic activity in myeloma cells	_	107, 108
TR3	Three consecutive extracellular TRAIL domains fused together in a head-to-tail configuration	Improved stability with activity similar to native TRAIL	RBC decorated TR3 (TR3-RBC) pancreatic cancer xenograft model	109
TRAIL-Mu3	Replacement of amino acid sequence VRERGPQR in TRAIL (114–121 aa) into RRRRRRR	Enhanced antitumour effects on pancreatic cancer cell lines	Pancreatic cancer xenograft model	110, 111
bvKD548-55	Bispecific death receptors 4 and 5 agonist from a kringle domain scaffold grafted with a loop	Advantage in triggering tumour cell death <i>via</i> DR5 and DR4 over TRAIL	-	113
APG350	Two single-chain TRAIL trimers dimerized by an Fc-part of human IgG1	Improved pharmacokinetics and enhanced antitumour activity	Colorectal cancer, pancreatic cancer and melanoma xenograft models	114, 115
ABBV-621	APG350 derivative containing IgG1-Fc point mutation N297S removing the glycosylation site	Improved pharmacokinetics and antitumour activity due to eliminated binding to Fcγ receptors and complement component C1q	Colorectal and pancreatic cancer xenograft models	116
HexaTR	TRAIL fused with molecular superglue SpyTag and SpyCatcher at N-terminus. Upon mixing together, SpyTag-TRAIL and SpyCatcher-TRAIL ligate to a hexameric TRAIL variant	Improved pharmacodynamics and significantly increased apoptosis induction in various cancer cells	20-50 times greater antitumour effect, resulting in eradication of several types of large tumour xenografts	117
ABD-HexaTR	ABD (albumin binding protein) fused to the N-terminus of HexaTR	Improved pharmacodynamics and significantly increased apoptosis induction in various cancer cells	20–50 times greater antitumour effect, resulting in eradication of several types of large tumour xenografts	117
SnHexaTR	Superglue tags SnoopTagJr/SnoopDogTag, individually fused at the N- or C-terminus of the TRAIL and dimerized by Snoopligase	Hexavalent SnHexaTR showed a longer serum half-life and greater tumour uptake	Eradication of 50% of COLO 205 xenograft tumours	118

#### Table 1 (continued).

Note. A dash indicates that no data on in vivo studies are available.

# 6. Bifunctional fusions of TRAIL with antitumour peptides or proteins

Genetically engineered bispecific proteins integrate functionally distinct protein fragments into a single molecule and thus exert diverse biological effects, such as acting as a drug or drug transporter, or both.<sup>119</sup> Fusions of TRAIL with antitumour peptides or proteins have shown promising potential to enhance its efficacy in cancer therapy. The conjugation of TRAIL with specific peptides or proteins can lead to improved cell recognition, targeted tumour delivery, and enhanced apoptotic signalling. As a part of these molecular constructs, TRAIL is able to selectively engage its cognate DRs on cancer cells, triggering the extrinsic apoptosis pathway while minimizing off-target effects on healthy tissues.<sup>120</sup> This strategy not only enhances the apoptotic response but also overcomes resistance mechanisms commonly found in cancer cells, ultimately increasing the overall therapeutic impact of TRAIL-based treatment in various malignancies.

Integrins  $\alpha\nu\beta3$  and  $\alpha\nu\beta5$  play crucial roles in various aspects of cancer progression and metastasis. Their upregulation is associated with tumour growth, angiogenesis and invasion, making them attractive targets for cancer therapy. Targeting integrins  $\alpha\nu\beta3$  and  $\alpha\nu\beta5$  has been explored as a potential strategy to inhibit cancer progression and metastasis, making them important targets for the development of anticancer therapeutics.<sup>121,122</sup> In particular, RGD (Arg-Gly-Asp) peptides specifically target the integrins  $\alpha\nu\beta3$  and  $\alpha\nu\beta5$ , which are commonly overexpressed in both the tumour neovasculature and tumour cells. RGD-L-TRAIL fusion protein combines the peptide ACDCRGDCFC, which contains the RGD cell adhesion motif, with TRAIL to enable double-targeted delivery of TRAIL to the tumour site. RGD-L-TRAIL showed promising results in inhibiting the proliferation of various cancer cell lines and significantly inhibiting tumour growth in mice with colon cancer tumour xenografts.<sup>123</sup> Another fusion RGD-TRAIL-NGR, containing the tumour-targeting peptides RGD and NGR at the N- and C-terminus of TRAIL, respectively, demonstrated improved anticancer efficacy and metastasis inhibition activity.<sup>124</sup> Another fusion protein, RGD-TRAIL-ELP, was developed by fusing the C-terminus of RGD-TRAIL with elastin-like polypeptides (ELPs), which were able to selfassemble into nanoparticles under physiological conditions. RGD-TRAIL-ELP demonstrated an enhanced apoptosisinducing ability, and a single intraperitoneal injection of nanoparticles resulted in almost complete tumour regression in the COLO-205 tumour xenograft model.<sup>125</sup>

RGD peptides can be either linear or cyclic. Cyclic RGD peptides have higher activity than linear RGD peptides due to their more stable conformation that resists proteolysis.<sup>126</sup> The cyclic iRGD peptide (CRGDKGPDC) exploits the tumour microenvironment by activating the integrin-dependent binding to the tumour vasculature, and neuropilin-1 (NRP-1)-dependent transport into tumour tissues.<sup>127</sup> A sTRAIL-iRGD fusion was developed to address the limitations of traditional TRAIL therapy, such as low tumour permeability and limited efficacy. The sTRAIL-iRGD achieved improved tumour-specific delivery, more effective apoptosis-inducing capacity compared to TRAIL, and exhibited antitumour effects more effectively in a gastric cancer model.<sup>128</sup> Another fusion peptide DR5-B-iRGD was generated by fusing iRGD to the C-terminus of the DR5selective TRAIL mutant variant DR5-B.129 DR5-B-iRGD penetrated into U87 tumour spheroids faster than DR5-B, demonstrated an enhanced antitumour effect in human glioblastoma cells and more effectively inhibited tumour growth in a xenograft mouse model of human glioblastoma.

The fusion of the tumour-homing RGR peptide (CRGRRST) with TRAIL to produce RGR-TRAIL has been reported.<sup>130</sup> RGR-TRAIL showed enhanced cell binding and cytotoxicity in colorectal cancer (CRC) cells compared to TRAIL, and exerted significantly enhanced growth suppression in mice bearing CRC tumour xenografts. The *in vitro* and *in vivo* antitumour effects of RGR-TRAIL were significantly improved by combination with (EGFR)-targeted photodynamic therapy (PDT).

The Fn14 (fibroblast growth factor-inducible 14-kDa protein) signalling axis, which is activated by its ligand TWEAK (TNF-like weak inducer of apoptosis), is involved in tumour cell proliferation through autocrine and paracrine signalling and can promote cancer progression through pro-survival and proangiogenic effects.<sup>131</sup> An Fn14-TRAIL fusion containing 1–79 amino acid fragment of the murine Fn14 receptor with murine TRAIL (118–291) was generated to attenuate an experimental autoimmune encephalomyelitis.<sup>132</sup> Later, the similar Fn14-TRAIL fusion of the extracellular domain of human Fn14 (1–52) with human TRAIL (53–217) demonstrated higher apoptotic activity than TRAIL alone in hepatocellular carcinoma cell lines and strong anticancer properties *in vivo*.<sup>133</sup>

Annexin V, a member of the Annexin superfamily, is known for its involvement in various cellular processes such as blood coagulation, signal transduction, and anti-inflammatory responses. In the context of cancer therapy, the development of a chimeric protein Annexin V-TRAIL (TP8) has shown promising results in the selective induction of apoptosis in various tumour cell types, both *in vitro* and *in vivo*.<sup>134</sup> This fusion protein exhibited superior efficacy compared to native TRAIL representing a potential strategy for the treatment of TRAIL-resistant cancers.

MUC16 (CA125), a well-established biomarker in ovarian cancer, is also highly expressed in other cancers including pancreatic and breast cancers. Meso-TR3, a fusion of TR3 (Ref. 109) with mesothelin, which specifically binds MUC16, is a novel therapeutic that induces potent cell death by targeting *via* an additional tumour-specific moiety. Meso-TR3 has been shown to have a significantly higher killing capacity on MUC16-expressing cancer cells compared to non-targeted TR3 and recombinant TRAIL.<sup>135</sup> The specificity of Meso-TR3 for MUC16-positive cells was validated in various *in vitro* and *in vivo* experiments, assuming its potential for homotypic (tumour cell-tumour cell) and heterotypic (tumour cell-mesothelial cell) cell interactions.<sup>136</sup>

The synthetic 5-amino acid peptide TMTP1 (NVVRQ) identified by the FliTrx bacterial peptide display system specifically binds highly metastatic tumour cells, including prostate, breast, lung and gastric cancers *in vitro* and *in vivo*, but not the non-metastatic cell lines.<sup>137</sup> In line with that, the sTRAIL-TMTP1 fusion protein demonstrated significant cytotoxic effects on highly metastatic tumour cells through the activation of the extrinsic apoptotic pathway. In preclinical studies, sTRAIL-TMTP1 effectively inhibited primary tumour growth and metastasis in various cancer models, and also showed a stronger suppressive effect on angiogenesis compared to standard TRAIL treatment.<sup>138</sup>

Second mitochondria-derived activator of caspase/direct inhibitor of apoptosis-binding protein with low pI (SMAC/DIABLO) is a pro-apoptotic mitochondrial protein that is released into the cytosol in response to various apoptotic stimuli and antagonizes inhibitors of apoptosis proteins (IAPs), thus allowing the activation of caspases and apoptosis. AD-053.2 is a

fusion protein in which an 8-amino acid fragment of the SMAC/ Diablo protein has been fused to the N-terminus of TRAIL with a metalloprotease cleavage linker.<sup>139</sup> The AD-053.2 protein showed a potent cytotoxic activity with the  $IC_{50}$  values in femtomolar range for the most sensitive cancer cell lines and significantly reduced the rate of the tumour growth in colon and lung adenocarcinoma animal models.

CD19, a B-lineage restricted receptor expressed on leukemia cells, represents a promising target for biotherapy in relapsed acute lymphoblastic leukemia due to its absence in critical nonhematopoietic organs and its abundant expression on relapsed rare mature B-cell leukemia clones. The recombinant human CD19L-sTRAIL fusion protein exhibited selective binding and potent apoptotic activity against CD19-positive leukemia cells, activating the TRAIL death pathway in a caspase 8-dependent manner. In preclinical studies, CD19L-sTRAIL demonstrated significant antileukemic potency against primary leukemia cells and in acute myeloid leukemia (AML) xenograft models without significant toxicity in animal models.<sup>140,141</sup> The favourable pharmacokinetic profile of the protein and efficacy in chemotherapy-resistant cases suggest its potential as a biotherapy for relapsed and refractory AML patients.

Lidamycin (LDM), an antitumour antibiotic, has potent cytotoxicity against cancer cells *in vitro* and *in vivo*.<sup>141</sup> The bifunctional Ec-LDP-TRAIL fusion protein combines the EGF C-loop (22 amino acids of the EGF C-terminus), the Lidamycin apoprotein (LDP) and the functional TRAIL. In addition, the energized fusion protein Ec-LDP-TRAIL-AE was created by mixing Ec-LDP-TRAIL with enediyne chromophore AE. Ec-LDP-TRAIL-AE exhibited specific binding to EGFR and TRAIL DRs and induction of apoptosis in tumour cells expressing these receptors. In both *in vitro* and *in vivo* studies, Ec-LDP-TRAIL-AE showed enhanced antitumour activity compared to LDM alone, while sparing normal organs such as the liver, lung, and intestine in experimental animals.<sup>142</sup>

The cytokine IL2 has been approved for the treatment of metastatic renal cell carcinoma and metastatic melanoma. It has been successfully used to combat various malignant neoplasms due to its ability to induce the proliferation of natural killer cells, T cells and B cells, thereby enhancing cellular immunity against cancer.<sup>143</sup> The IL2-TRAIL fusion showed higher cytotoxicity in leukemic cell lines compared to TRAIL. This immunotoxin was found to be highly specific in targeting CD25-positive leukemia cells, showing the potential as a potent and selective therapeutic agent.<sup>144</sup>

Antimicrobial peptides (AMPs) with high efficacy and low toxicity are promising new drugs that can replace chemoradiotherapy.<sup>145</sup> The antibacterial peptide CM4 is a cationic linear  $\alpha$ -helical 35-amino acid peptide, which belongs to the cecropin family. The most intriguing feature of CM4 is its destructive effect on bacteria, fungi and tumours without damaging normal cells. TRAIL-CM4, a fusion protein of TRAIL and CM4, has shown potent antitumour activity, inducing higher rates of cell death and apoptosis in cancer cells compared to TRAIL alone.<sup>146</sup>

sTRAIL-melittin<sup>147</sup> is another TRAIL fusion with a 26-amino acid antibacterial peptide melittin with anticancer activity, derived from bee venom.<sup>148</sup> Despite its potential to induce tumour cell death through multiple pathways, the non-selective toxicity and side effects of melittin restrict its application as a viable anticancer agent *in vivo*. sTRAIL-melittin fusion displayed enhanced anticancer activity against leukemia and liver cancer cells, inducing significant apoptosis, while demonstrating no cytotoxic effects on normal erythrocytes and human embryonic kidney cells,<sup>147</sup> highlighting its potential for cancer therapy.

The selective apoptosis activator Par-4 (prostate apoptosis response-4) is a multi-domain protein containing the 59-amino acid SAC (selective for apoptosis of cancer cells) domain, which enables it to specifically induce apoptosis in cancer cells without harming normal cells or tissues.<sup>149</sup> Fusing the Par-4 SAC domain with TRAIL resulted in enhanced anticancer potential of the resulting SAC-TRAIL fusion, creating a promising candidate for cancer therapy. The structure and sequence order of the fusion protein were optimized using various flexible linkers for expression in *E. coli*.<sup>150</sup>

Enhancement of the anticancer effect of TRAIL was demonstrated by a TGF3L-TRAIL fusion, in which a synthetic TGF3L (the third disulfide loop of TGF- $\alpha$ ) peptide was linearized to eliminate any possible residual EGFR binding affinity and fused to the N-terminus of TRAIL.<sup>151</sup> The TGF3L-TRAIL fusion showed increased cytotoxicity in a variety of cancer cell lines in vitro and in vivo. The authors showed that TGF3L-TRAIL forms stable polymers, which provides increased cytotoxicity against cancer cells. Importantly, the TGF3L-TRAIL fusion was designed with a His tag at the N-terminus. In another attempt, the truncated sequence from the N-terminus of hCtr1 (human copper transporter 1) (NCTR25tag) was used instead of the classic His-tag, which is not well suited for clinical applications to produce TRAIL in E. coli.<sup>151,152</sup> Both NCTR<sub>25</sub>-TRAIL and NCTR<sub>25</sub>-TGF3L-TRAIL demonstrated the ability to self-organize into a polymerlike structures, and exhibited significantly higher activity compared to TRAIL via selective activation of DR4 and DR5 receptors.

To improve the penetration of TRAIL molecules across the blood-brain barrier (BBB), the small 19-amino acid peptide ANG2 (Angiopep-2) was fused to single-chain scTRAIL via the crystallizable fragment (Fc) domain of a human IgG including the hinge region for the treatment of glioblastoma (Fig. 4).<sup>153</sup> The Angiopep family of peptides are derived from the Kunitz domain of human aprotinin, which can cross the BBB and are used to facilitate the delivery of pharmacological agents to the brain. The Angiopep-2 has demonstrated the ability to facilitate LRP1 (low-density lipoprotein receptor-related protein 1)-dependent transcytosis across the BBB and has been used to enhance the CNS (central nervous system) penetrance of various cargoes, including drugs, proteins and nanoparticlebased systems, with early clinical trials showing promisingly low toxicity.154 The hexavalent scTRAIL-Fc-ANG2 fusion protein remained highly effective in inducing apoptosis in glioblastoma cells, but binding to BBB cells was predominantly due to the TRAIL protein as TRAIL has a significantly higher binding rate for its receptors than ANG2 to the LRP1 receptor.<sup>153</sup>

The formation of new blood vessels (angiogenesis) is a complex and dynamic process regulated by various pro- and antiangiogenic molecules. Angiogenesis plays a key role in tumour growth, invasion and metastasis.<sup>155</sup> Treatment of tumour diseases with antiangiogenic agents is a promising strategy for antitumour therapy. However, the use of these drugs is still limited due to several disadvantages such as side effects, acquired drug resistance and tumour recurrence. In order to simultaneously target the tumour and tumour microenvironment, several TRAIL fusions with antiangiogenic peptides have been developed.

Vasostatin is a naturally occurring endogenous peptide derived from the cleavage of calreticulin, a multifunctional protein involved in several cellular processes. This peptide has



**Figure 4.** Design of a CNS-targeted TRAIL-receptor agonist. (*a*) Functional units, (*b*) composition and (*c*) schematic assembly of CNS-targeted scTRAIL variants and relevant control proteins. Reproduced from Moorthy *et al.*<sup>153</sup> under the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

been implicated in the regulation of angiogenesis, the process by which new blood vessels are formed from the pre-existing ones.<sup>156</sup> VAS-TRAIL, a fusion protein combining vasostatin and TRAIL moieties, inhibited tumour cell growth and induced apoptosis in cancer cells, while significantly suppressing proliferation in fetal bovine heart endothelial (FBHE) cells. The combined anti-angiogenic properties of vasostatin and apoptotic effects of TRAIL resulted in increased endothelial cell apoptosis compared to either vasostatin or sTRAIL alone, demonstrating the effective integration of both domains within the VAS-TRAIL fusion protein.<sup>157</sup>

Another fusion protein, AD-O51.4, is composed of a TRAILderived sequence linked to VEGFA-derived peptides. AD-O51.4 engages a multifaceted mechanism, involving both extrinsic and intrinsic apoptotic pathways, and triggers potent inhibition of tumour growth. Its potential to induce apoptosis in both tumour cells and endothelial cells highlights the importance of targeting the tumour microenvironment for effective therapeutic intervention.<sup>158</sup>

AD-O51.4 reduced the growth of CRC PDXs (patientderived xenografts) with good efficacy and displayed cytotoxic effects in DLBCL (diffuse large B-cell lymphoma) cells through induction of DR-mediated caspase-dependent apoptosis.<sup>159,160</sup>



**Figure 5.** Fusion of DR5-selective TRAIL mutant variant **DR5-B** with antiangiogenic synthetic peptide **SRH** (SRHTKQRHTALH) and cell-penetrating peptide iRGD (CRGDKGPDC) for dual targeting of VEGFR2 and integrin  $\alpha\nu\beta3$  receptors.

We have also recently generated TRAIL-based fusions **SRH-DR5-B**<sup>161</sup> and **SRH-DR5-B-iRGD**<sup>162</sup> with antiangiogenic synthetic peptide **SRH** (SRHTKQRHTALH) at the N-terminus of DR5-selective TRAIL mutant variant **DR5-B**<sup>33</sup> or **DR5-B-iRGD**<sup>129</sup> *via* flexible linker (Fig. 5). **DR5-B** selectively binds the DR5 receptor and induces apoptosis more potently than TRAIL.<sup>13,61,163</sup> Both **DR5-B**-based fusions demonstrated enhanced antitumour activity *in vitro* and *in vivo* compared to **DR5-B**. Importantly, they internalized the DR5 receptor faster than **DR5-B** and exhibited antiangiogenic activity in glioblastoma xenografts, with these effects being more pronounced for **SRH-DR5-B-iRGD** due to the additional  $\alpha\nu\beta3$  integrin-specific iRGD peptide moiety.

A summary of TRAIL-based bifunctional fusions is presented in Table 2.

### 7. TRAIL fusions with antibody fragments targeting tumour-specific antigens

Single-chain fragment variable (scFv) antibodies have become valuable tools in cancer therapy due to their ability to specifically target tumour-associated antigens. These are single 25 kDa polypeptides that contain the variable light chain (VL) and the variable heavy chain (VH) regions of the antibody connected by a flexible linker peptide, which is usually 15–20 amino acids long and consists of glycine and serine with dispersed hydrophilic residues for increased solubility.<sup>164,165</sup> Single-chain variable fragments, which contain the complete antigen-binding domains of the whole antibody, have several advantages, such as high specificity and affinity for antigens, low immunogenicity, and the ability to penetrate and diffuse into tumour tissue.<sup>166</sup>

Over the past decades, numerous TRAIL or single-chain TRAIL (scTRAIL) fusions with scFc antibodies against various target surface antigens known to be highly expressed by certain tumour types have been generated to enhance

12 of 27
----------

Name	Structure	Effects	In vivo model	Ref.
RGD-L-TRAIL	TRAIL fusion with ACDCRGDCFC peptide	Enhanced apoptotic activity, effective tumour growth suppression	Xenograft model of colorectal cancer	123
RGD-TRAIL-NGR	TRAIL fusion with RGD and NGR peptides at the N- and C-termini	Potent anti-metastasis effects <i>in vitro</i> and <i>in vivo</i>	Xenograft model of human fibrosarcoma	124
RGD-TRAIL-ELP	TRAIL fusion with RGD peptide and ELP (elastin-like polypeptides), self- assembled into nanoparticle	Significant tumour cell apoptosis without apparent liver toxicity	Tumour regression in the COLO-205 xenograft model	125
sTRAIL-iRGD	TRAIL fusion with iRGD (CRGDKGPDC) peptide	Enhanced tumour penetration and antitumour activity	Xenograft model of human gastric carcinoma	128
DR5-B-iRGD	DR5-selective TRAIL variant <b>DR5-B</b> fused with iRGD peptide	Enhanced penetration and antitumour activity	Xenograft model of human glioblastoma	129
RGR-TRAIL	Tumour-homing peptide RGR (CRGRRST) fusion with TRAIL	Enhanced tumour penetration and antitumour activity	Xenograft model of colorectal cancer	130
Fn14-TRAIL	Murine Fn14 (1-79) fused to murine TRAIL (118-291)	Attenuated experimental auto- immune encephalomyelitis	Chronic paralytic disease model in C57BL/6 mice	132
Fn14-TRAIL	Human Fn14 (1-52) fused to human TRAIL (53-217)	Enhanced anticancer potential	Xenograft model of hepatocellular carcinoma	133
TP8	TRAIL fusion with Annexin V	Higher efficacy than TRAIL alone <i>in vitro</i> and <i>in vivo</i>	Xenograft models of colo- rectal, breast and hepatocellular carcinoma	134
Meso-TR3	TRAIL trimer TR3 fusion with soluble mesothelin	Efficient binding and killing of MUC16- expressing tumour cells	Xenograft mouse model of ovarian cancer	135, 136
sTRAIL-TMTP1	TRAIL fusion with TMTP1 (tumour molecular targeted peptide 1) NVVRQ	Enhanced antitumour activity against highly metastatic cancers	Xenograft mouse models of human prostate cancer	138
AD-053.2	8-amino acid fragment of the Smac/Diablo fused to N-terminus of TRAIL with a metalloprotease cleavage linker	Enhanced cytotoxicity in various cancer cell lines and significant reduction of the tumour growth rate	Colon and lung adeno- carcinoma animal models	139
CD19L-sTRAIL	Natural ligand of the human CD19 receptor fused to sTRAIL with a flexible linker	More favourable pharmacokinetic profile than sTRAIL, significant antileukemic potency	Leukemia xenograft models in NOD/SCID mice	140
Ec-LDP-TRAIL	22-amino acid C-loop of EGF and apoprotein (LDP) of antitumour antibiotic lidamycin fused to N-terminus of TRAIL	Potent and selective cytotoxicity against different carcinoma cells	Human epidermoid carcinoma xenografts	142
IL2-TRAIL	IL2 fused to TRAIL via a Gly <sub>3</sub> Ser linker	Efficient killing of myeloid and lymphoid leukemic patient cells	-	144
TRAIL-CM4	TRAIL fusion with antibacterial peptide CM4 via a $(Gly_4Ser)_3$ linker	Enhanced apoptotic activity in leukemia cells	-	146
sTRAIL-melittin	Melittin fusion with the N-terminus of sTRAIL <i>via</i> a $(Gly_4Ser)_3$ linker	Apoptotic activity in leukemia cells and liver carcinoma cells	-	146
SAC-TRAIL	TRAIL fusion with SAC (selective for apoptosis core domain) of Par-4 protein <i>via</i> a linker peptide $(Gly_4Ser)_3$	Improved targeting ability, cytotoxic effects and apoptosis induction	Xenograft model of ovarian carcinoma and breast cancer	150
His-TGF3L-TRAIL NCTR <sub>25</sub> -TGF3L- TRAIL	TGF3L (the third disulfide loop of TGF- $\alpha$ fusion) peptide fused to the N-terminus of TRAIL with His- or NCTR <sub>25</sub> tags	Assembly into complexes, increased antitumour activity	Lung and colon cancer xenografts	151, 152
scTRAIL-Fc-ANG2	19-mer synthetic BBB-shuttle peptide ANG2 (Angiopep-2) fused to the C-ter- minus of single-chain trimeric TRAIL <i>via</i> the crystallisable fragment (Fc) domain of a human IgG including the hinge region	Similar antitumour activity as Fc-fused single chain TRAIL (Fc-scTRAIL) in cancer cells	-	153
VAS-TRAIL	Vasostatin (120–180 aa domain) fused to N-terminus of TRAIL <i>via</i> a linker peptide	Apoptotic activities toward lung cancer and heart vascular endothelium cells	_	157
AD-051.4	TRAIL N-terminal fusion with positively charged VEGFA-derived effector peptides	Increased half-life and apoptotic activity in multiple cancer cell lines	Potent antitumour activity multiple cancer mouse xenografts	158, 160
SRH-DR5-B	DR5-selective TRAIL variant <b>DR5-B</b> N-terminal fusion with synthetic antiangiogenic peptide <b>SRH</b>	Enhanced cytotoxic activity in 2D and 3D models of multiple cancer cell lines	_	161
SRH-DR5-B-iRGD	DR5-selective TRAIL variant <b>DR5-B</b> fusion with synthetic anti-angiogenic peptide <b>SRH</b> and iRGD at the N- and C-termini	Improved anticancer activity in glioblastoma, <i>in vivo</i> antiangiogenic activity	Xenograft model of human glioblastoma	162
NT / A 1 1 1 1		1		

 Table 2. TRAIL fusions with peptides or full-length functional proteins.

Note. A dash indicates that no data on in vivo studies are available.

antitumour activity and improve pharmacokinetic parameters.<sup>19,20,167</sup>

The first scFv-Fc fusion, MBOS4-TRAIL, was generated in 2001 by fusing a recombinant tumour stroma marker FAP (fibroblast activation protein)-specific single-chain antibody fragment MBOS4 and the Fc fragment of human IgG to the extracellular domain of TRAIL *via* linkers. Dose-response analysis of MBOS4-TRAIL-induced apoptosis revealed approximately 30-fold higher toxicity towards FAP-expressing cells compared to TRAIL.<sup>168</sup>

Two fusions with antitumour bystander activity, scFvC54:sTRAIL and scFvCD7:sTRAIL, were generated by fusing the scFv fragments specific for epithelial glycoprotein 2 (C54) and cell surface glycoprotein (CD7) to the N-terminus of human sTRAIL. The scFvC54:sTRAIL showed favourable properties, potentially reducing the amount of sTRAIL required for antitumour activity.<sup>169,170</sup> The scFvCD7:sTRAIL, which contains an scFv antibody fragment, specific for human CD7, which is abundantly expressed on most T cell malignancies and 10% of acute myeloid leukemias, showed enhanced and target antigen-restricted apoptotic activity against human T cell acute lymphoblastic leukemia (T-ALL) cells without toxicity to normal human blood and endothelial cells. In mixed culture experiments with CD7-positive and CD7-negative tumour cells, scFvCD7:sTRAIL induced very potent bystander apoptosis.<sup>171</sup>

The EGFR signalling plays a key role in the regulation of various tumour cell functions such as cell cycle progression, inhibition of apoptosis, angiogenesis, cell motility, adhesion, and invasion.<sup>172</sup> An engineered EGFR-specific scFv425:sTRAIL fusion protein constructed of a fragment of the EGFR-blocking antibody scFv425 genetically fused with soluble TRAIL, demonstrated a significant apoptosis-inducing activity in a number of EGFR-positive tumour cell lines.<sup>173</sup> Treatment of mice bearing intraperitoneal renal cell carcinoma xenografts with the adenovirus-expressed Ad-scFv425:sTRAIL fusion resulted in a rapid and massive reduction in tumour burden and subsequent long-term survival.<sup>174</sup>

A pan-B cell marker CD19 has been recognized as a potential immunotherapy target for B cell disorders, including bloodborne malignancies and autoimmune diseases.<sup>175</sup> The scFvCD19:sTRAIL fusion protein demonstrated CD19-specific apoptosis induction in B-ALL cell lines, while sparing normal hematopoietic cells. It also showed bystander apoptosisinducing effects in CD19-negative tumour cells and significant therapeutic potential in inhibiting tumour engraftment and prolonging survival in mice xenografted with human B-ALL cells.<sup>176</sup>

Another antigen-specific antibody fragment against CD33 was used to generate the scFvCD33:sTRAIL fusion.<sup>177</sup> CD33 is a cell surface antigen expressed in  $\sim 80-90\%$  of AML patients, particularly on leukemic blasts, making it a target for therapy.<sup>178</sup> *Ex vivo* treatment of patient-derived CD33-positive AML cells with scFvCD33:sTRAIL resulted in potent induction of apoptosis. In chronic myeloid leukemia (CML) cells, scFvCD33:sTRAIL had potent antileukemic activity against CD33+ cells when combined with the Bcr-Abl tyrosine kinase inhibitor, imatinib mesylate (Gleevec).<sup>177</sup>

The anti-MCSP:sTRAIL fusion protein designed to target melanoma cells with overexpressed CSPG4 (chondroitin sulfate proteoglycan 4), demonstrated potent apoptosis induction and tumour growth retardation in preclinical models.<sup>179</sup> CSPG4 is overexpressed in several types of cancer, including breast cancer, melanoma, squamous cell carcinoma, mesothelioma, neuroblastoma, adult and pediatric sarcomas, and some

hematological cancers. CSPG4 has been the target of numerous anticancer therapies aimed at inhibiting its signalling pathways that promote proliferation, migration, and invasion.<sup>180</sup> Targeting MCSP with anti-MCSP:TRAIL inhibited MCSP signalling and activated TRAIL apoptotic signalling in melanoma cells *in vitro* and *in vivo*.<sup>181</sup>

To augment the tumouricidal activity of T cells, two recombinant fusion proteins, anti-CD3:TRAIL and K12:TRAIL, were generated that selectively bind to the surface of T cells with CD3 and CD7 antigens, respectively. Anti-CD3:TRAIL contains a CD3 stimulatory antibody fragment, whereas K12:TRAIL contains a soluble form of the CD7 ligand K12. Both fusions strongly enhanced the tumouricidal activity of T cells against a panel of cancer cell lines, primary patient-derived malignant cells, and in a murine xenograft model.<sup>182</sup>

The voltage-gated potassium channel Kv10.1, a tumourspecific marker, has been recognized as a promising target in cancer therapy due to its high expression in tumour tissues.<sup>183</sup> A KV10.1-specific scFv antibody fused with sTRAIL has been developed for combination therapies to target prostate cancer cells with high tumour specificity.<sup>184</sup> The scFv62-TRAIL construct induced apoptosis in human prostate cancer cells after sensitization with cytotoxic drugs only in KV10.1-positive cancer cells, but not in non-transformed cells. The antitumour effect of scFv62-TRAIL in combination with doxorubicin was demonstrated in SCID mice bearing subcutaneous melanomas.<sup>185</sup>

CD47 is a key 'don't eat me' signalling molecule that allows cancer cells to evade phagocytic clearance. Binding of CD47 to a SIRP $\alpha$  receptor on the surface of phagocytes inhibits the cancer cell clearance. Blocking the CD47-SIRP $\alpha$  interaction can enhance phagocytic clearance of cancer cells, making it a potential therapeutic approach to boost the efficacy of anticancer antibodies and promote apoptosis in malignant cells.<sup>186</sup> Anti-CD47:TRAIL fusion enhanced rituximab (RTX)-mediated phagocytosis of B cell non-Hodgkin's lymphoma (B-NHL) cells and triggered apoptosis in CD47+ B cell lines and primary malignant B-NHL samples while sparing normal blood cells.<sup>187</sup>

Selective expression of C-type lectin-like molecule-1 (CLL1) on granulocytes, monocytes, and dendritic cells makes it a promising target for enhancing the antitumour activity of granulocytes for the AML therapy.<sup>188</sup> CLL1:TRAIL is a fusion protein designed to enhance the ability of leukocytes, primarily granulocytes, to significantly enhance their antitumour activity and increase the cytotoxicity of therapeutic anticancer antibodies. CLL1:TRAIL improved the overall efficacy of anti-cancer antibodies such as rituximab, cetuximab, alemtuzumab and daratumumab-based antibodies.<sup>189</sup>

Based on the Flag-TNC-hTRAIL fusion,108 two novel proteins, scFv:G28-TRAIL and scFv:lahCD70-TNC-TRAIL, bearing CD40- and CD70-specific scFv fragments have been constructed. CD40, a member of the TNF receptor superfamily, has shown promise in cancer immunotherapy by promoting antitumour immune responses when activated. Clinical trials using CD40 agonist antibodies, often in combination with other treatments such as checkpoint inhibitors and chemotherapy, have demonstrated encouraging antitumour effects, making CD40 activation a valuable strategy for improving cancer treatment.<sup>190</sup> CD40-dependent enhancement of apoptosis by scFv:G28-TRAIL has been demonstrated in various cell lines. providing potential for CD40-restricted cancer therapy.<sup>191</sup> Another member of the TNF family, CD70, interacts with the CD27 receptor to promote the expression of anti-apoptotic genes that boost T cell survival and proliferation.<sup>192</sup> The scFv:lahCD70-TNC-TRAIL fusion, a stabilized form of TRAIL

containing a llama CD70-specific scFv, demonstrated enhanced induction of apoptosis upon CD70 binding through effective interference with the CD70-CD27 interaction. scFv:lahCD70-TNC-TRAIL fusions containing DR4- and DR5-selective TRAIL mutant variants also enhanced the induction of cell death upon CD70 binding, with a preference for the DR5-selective variant.<sup>193</sup>

The scFvCD20-sTRAIL fusion was designed and expressed in human umbilical cord mesenchymal stem cells as a carrier for dual targeted therapy against non-Hodgkin's lymphoma.<sup>194</sup> The expression of non-glycosylated CD20 protein has been detected on the surface of normal and malignant B cells and is considered an ideal therapeutic target, as rituximab-based immunotherapy has become the standard of care for most B-cell malignancies.<sup>195</sup> The scFvCD20-sTRAIL fusion protein demonstrated significant enhancement of cellular apoptosis through both extrinsic and intrinsic apoptotic signalling pathways. In the NOD/SCID mouse BJAB subcutaneous lymphoma xenograft model, intravenous injection of MSC.scFvCD20-sTRAIL significantly inhibited tumour growth compared to MSC.ISZ-sTRAIL-treated mice.<sup>194</sup>

MRP3, a multidrug resistance protein, has been identified as a specific antigen in glioblastoma multiforme (GBM), making it a valuable target for antibody-based cancer therapy.<sup>196</sup> A scFvM58-sTRAIL fusion protein, designed to selectively induce apoptosis in MRP3-positive GBM cells, demonstrated promising target antigen-restricted pro-apoptotic activity, providing a potential therapeutic approach for GBM.<sup>197</sup>

Desmoglein-3 (DSG3) has been characterized as a key mediator involved in desmosome remodelling, epidermal proliferation and differentiation, cell migration, and apoptosis, indicating that DSG3 plays a major role in tissue integrity and homeostasis. Depletion of DSG3 in HaCaT keratinocytes resulted in suppression of cell proliferation and colony growth.<sup>198</sup> Px44-TRAIL fusion was constructed using the anti-desmoglein mAb Px44 to target keratinocytes which express DSG3 on their surface. This fusion was biologically active and stable, and was successfully delivered to keratinocytes.<sup>199</sup>

Another antitumour antibody fragment with specificity for PD-L1 (programmed death ligand 1) was used to generate the anti-PD-L1:TRAIL fusion protein. PD-L1 is a key immune regulator in cancer immunotherapy, and antibodies against PD-L1 or its receptor PD-1 prevent PD-L1-mediated inhibition of antitumour T cells, thereby promoting antitumour immunity.<sup>200</sup> In experiments with mixed T cells and cancer cell cultures, anti-PD-L1:TRAIL fusion increased T cell activation and IFN $\gamma$  secretion, resulting in increased destruction of cancer cells of various origins, including primary patient-derived cancer cells. Interestingly, anti-PD-L1:TRAIL converted immunosuppressive PD-L1-expressing myeloid cells into proapoptotic effector cells that triggered TRAIL-mediated cancer cell death.<sup>201</sup>

A bifunctional ENb-TRAIL fusion of the EGFR-targeting nanobody ENb derived from heavy chain-only antibodies found in camelids, and TRAIL ligand showed therapeutic efficacy in different cancer cell types unresponsive to either EGFR antagonist or TRAIL DRs agonist monotherapies. In a mouse model of primary glioblastoma, engineered stem cells (SCs) expressing ENb-TRAIL encapsulated in a synthetic extracellular matrix (SCENb-TRAIL) alleviated tumour burden and significantly prolonged survival.<sup>202</sup>

Platelet-derived growth factor receptor beta (PDGFR $\beta$ ) is a receptor protein that is often overexpressed in tumour-associated pericytes, suggesting its potential as a target for tumour therapy.

When PDGFRs are inhibited, cancer growth, metastasis, invasion and angiogenesis are reduced, enhancing the antitumour effects of cancer treatments.<sup>203</sup> TRAIL fusion with the anti-PDGFRß affibody Z-hTRAIL mediated PDGFRß-dependent binding of hTRAIL to pericytes and killed tumour cells through juxtatropic activity, or exhibited cytotoxicity in tumour cells after release from pericytes. Fusion with the anti-ZPDGFRß affibody increased tumour uptake of hTRAIL, thereby enhancing the antitumour effect of hTRAIL in mice bearing colon carcinoma tumour xenografts without increasing acute liver and kidney toxicity.204 Compared to hTRAIL, Z-hTRAIL showed greater in vitro cell binding and apoptosis induction in aHSCs (hepatic stellate cells). In vivo experiments showed that the antihepatofibrotic effect of hTRAIL was improved by PDGFRβtargeted delivery.<sup>205</sup> PEGylation of Z-hTRAIL with 10 kDa PEG (polyethylene glycol) improved its pharmacokinetics and was more effective than Z-hTRAIL in resolving liver fibrosis. To improve the tumour-targeting ability and pharmacokinetics of TRAIL, a tridomain Z-ABD-TRAIL fusion protein was developed by fusing the tumour-homing ZPDGFR<sup>β</sup> affibody and an albumin-binding domain (ABD) to the N-terminus of TRAIL. The tridomain TRAIL variant exhibited increased tumour uptake and antitumour effect in mice bearing COLO205 tumour xenografts.<sup>206</sup>

Members of the epidermal growth factor receptor (HER), EGFR (ERB1) and four closely related receptor tyrosine kinases receptors HER2 (ErbB2), HER3 (ErbB3), and HER4 (ErbB4) are overexpressed in various human cancers, exhibiting tyrosine kinase-dependent oncogenic activity. Numerous tyrosine kinase inhibitors and monoclonal antibodies have been approved for clinical use.<sup>207</sup> EGFR is both a major oncogenic driver and a therapeutic target, however current EGFR inhibitors cause cancer resistance and this remains an important unmet clinical problem. To enhance apoptotic activity, several tumourtargeting scTRAIL fusion proteins with scFv fragments specific to various HERs have been developed. ErbB2-targeting fusion scFv-scTRAIL was found to exhibit superior efficacy both in vitro and in vivo compared to non-targeted TRAIL, highlighting its potential as a promising strategy for ErbB2targeted cancer treatment.<sup>208</sup>

A humanized scFv huC225 derived from cetuximab was fused to the N-terminus of scTRAIL, forming a scFva<sub>EGER</sub>scTRAIL fusion, with dimerization achieved by reducing the linker between VH and VL to form a  $Db\alpha_{EGFR}$ -scTRAIL fusion.209,210 Cancer cell lines required sensitization with either cycloheximide or bortezomib to elicit efficient TRAIL-mediated induction of apoptosis in vitro by these fusions, and dimerization of the fusion enhanced the induction of apoptosis. However, in the in vivo tumour model, DbaEGFR-scTRAIL demonstrated potent antitumour activity even in the absence of chemotherapeutic agents. In addition, the dimeric tetravalent fusion protein scFv-EHD2-scTRAIL has been developed by fusing the humanized anti-EGFR scFv to the N-terminus of dimeric EHD2-scTRAIL fusion containing IgE heavy chain domain 2 (EHD2).<sup>211</sup> The scFv-EHD2-scTRAIL fusion was more effective in killing tumour cell and showed higher antitumour activity in murine xenograft models than EHD2scTRAIL. Later, Fc-scTRAIL and scFc-Fc-scTRAIL fusions with enhanced antitumour activity were developed by replacing EHD2 IgGE with human IgG1 Fc.<sup>212</sup> Based on the Fc-scTRAIL, novel fusions were generated with scFv fragments specific for the HER2 (scFv4D5-Fc-scTRAIL) and HER3 (scFv3M6-FcscTRAIL and scFv323/A3hu3-Fc-scTRAIL) receptors, as well as the scFv323/A3hu3-Fc-scTRAIL fusion with scFv to

EpCAM (epithelial cell adhesion molecule).<sup>213</sup> All fusions demonstrated the comparable apoptotic activity in colorectal cancer cell lines. However, in xenograft tumour model, strongest antitumour activity was observed for the anti-HER3 scFv-Fc-scTRAIL fusion with complete tumour remissions. At the same time, untagged fusion Fc-scTRAIL showed a similar *in vivo* activity.

CD38 has been identified as a potential target for therapeutic antibodies in the treatment of multiple myeloma, with antibody products such as daratumumab showing encouraging results in clinical trials both alone and in combination with various chemotherapies, suggesting its efficacy as a viable treatment option.<sup>214</sup> A novel dual cytokine–antibody fusion IL2- $\alpha$ CD38- $\alpha$ CD38-scTRAIL, designed to target CD38-positive multiple myeloma (MM) cells, exhibited exceptional biochemical properties with retained binding specificity to CD38 and a potent ability to induce selective cell death *in vitro*.<sup>215</sup>

To address the limitations of previous treatments, two fusions, scFv62-scTRAIL and VHH-D9-scTRAIL, with

specificity to the voltage-gated potassium channel Kv10.1 based on the previously designed construct <sup>184</sup> were described. Fusion with newly generated VHH-D9 nanobody demonstrated higher antigen affinity and rapid and potent induction of apoptosis in various cell culture tumour models.<sup>216</sup>

A novel Fc-engineered CD19-targeting IgG1 antibody fused to a single-chain TRAIL domain, CD19-TRAIL, was developed as a new potential immunotherapeutic agent against acute lymphoblastic leukemia (BCP-ALL). This fusion demonstrated selective binding capacity and strong induction of apoptosis in CD19-positive BCP-ALL cell lines *in vitro* and *in vivo*, and significantly prolonged the survival of mice transplanted with BCP-ALL patient-derived xenografts with different cytogenetic backgrounds, especially when combined with the BCL-2 inhibitor venetoclax.<sup>217</sup>

A summary of TRAIL fusions with antibody fragments targeting tumour-specific antigens is presented in Table 3.

Table 3.	TRAIL fusions with antibody fragments targeting tumour-specific antigens.

Name	Target antigen	Effects	In vivo activity	Ref.
		scFv-TRAIL		
MBOS4-TRAIL	FAP (fibroblast activation protein)	Enhanced cytotoxicity in FAP expressing cells	-	168
scFvC54:sTRAIL	EGP2 (epithelial glycoprotein 2)	High cytotoxicity in leukemia and glioblastoma cell lines	-	169, 170
scFvCD7:sTRAIL	CD7 (cell surface glycoprotein)	Activity against T-cell leukemia	-	171
scFv425:sTRAIL	Epidermal growth factor receptor (EGFR)	Potent apoptosis induction in EGFR-positive tumour cell lines	Massive tumour reduction, long-term survival of renal cell carcinoma xenografts	173, 174
scFvCD19:sTRAIL	CD19B (lymphocyte antigen)	Potent apoptosis in several CD19-positive tumour cell lines	Prevention of Nalm-6 xeno- transplanted cells engraftment in mice	176
scFvCD33:sTRAIL	CD33 (myeloid cell surface antigen)	Potent anti-leukemia activity towards CD33 positive AML cells in combination with tyrosine kinase inhibitors	-	177
anti-MCSP:sTRAIL	MCSP (melanoma chondroitin sulfate proteoglycan)	Inhibition of MCSP signaling and activation of TRAIL apoptotic signalling in melanoma cells	Significant growth retardation of established melanoma xenografts	179
anti-CD3:TRAIL	CD3 (cluster of differentiation 3)	Strong inhibition of tumour growth and increased survival of xenografted mice by T cell surface delivery of TRAIL	Antitumour activity in colorectal carcinoma bearing mice	182
scFv62-TRAIL	KV10.1 (voltage-gated potassium channel)	Apoptosis induction in KV10.1-positive cancer cells	Antitumour activity with doxorubicin in melanoma xenografts	184, 185
Anti-CD47:TRAIL	CD47 (cluster of differentiation 47)	Enhancement of rituximab-mediated phagocytosis and triggering of CD47-restricted apoptosis in malignant B cells	-	187
CLL1:TRAIL	CLL1 (C type lectin-like molecule-1)	Augmenting tumouricidal activity of granulocytes, particularly when combined with therapeutic antibodies	_	189
scFv:G28-TRAIL	CD40 (TNF receptor superfamily member 5)	Enhanced TRAIL death receptor signaling in CD40-expressing cells	-	191
scFv:lαhCD70-TNC- TRAIL	CD70 (cluster of differentiation 70)	Strongly enhanced apoptosis induction upon CD70 binding in multiple cancer cells	-	193
scFvCD20-sTRAIL	CD20 (B-lymphocyte antigen)	Potent inhibition of cell proliferation in CD20-positive lymphoma cells	Significant inhibition of tumour growth in mice when compared MSC.ISZ-sTRAIL	194
scFvM58-sTRAIL	MRP3 (multidrug re- sistance protein 3)	High apoptotic activity in MRP3-expressing glioblastoma cell lines	-	197

Name	Target antigen	Effects	In vivo activity	Ref.
		scFv-TRAIL		
Px44-TRAIL	Dsg3 (desmoglein)	Apoptosis of the hyperproliferative, but not differentiating, cultured keratinocytes	_	199
anti-PD-L1:TRAIL	PD-L1 (programmed death-ligand 1)	PD-L1-directed TRAIL-mediated cancer cell death in PD-L1-expressing cells	-	201
ENb-TRAIL	EGFR (epidermal growth factor receptor)	Therapeutic efficacy in tumour cells from different cancer types	Alleviated tumour burden and increased survival of mice bearing glioblastoma tumour xenografts	202
Z-hTRAIL	PDGFR $\beta$ (platelet-derived growth factor receptor $\beta$ )	Improved tumour uptake and antitumour efficacy	Enhanced antitumour effect on colorectal cancer xenografts in mice	204
Z-hTRAIL-10K (PEGylated TRAIL)	PDGFRβ (platelet-derived growth factor receptor beta)	Regression of liver fibrosis more potent than with Z-hTRAIL	-	205
Z-ABD-TRAIL	PDGFRβ, Albumin	Improved pharmacokinetics and stability with activity similar to native TRAIL	Increased tumour uptake and antitumour effect on colorectal cancer xenografts in mice	206
	Sing	ele chain TRAIL trimer (scTRAIL) derivatives		
scFv-scTRAIL	ErbB2 (human epidermal growth factor receptor 2)	Higher therapeutic activity <i>in vitro</i> and <i>in vivo</i> than scTRAIL	Antitumour activity on colorectal cancer xenografts in mice	208
$Db\alpha_{EGFR}$ -scTRAIL	EGFR	Increased activity EGFR-positive or negative cancers cell lines	Superior antitumour activity on colorectal cancer xenografts in mice	209, 210
scFv-EHD2-scTRAIL	EGFR	8- to 18-fold increased cytotoxic activity compared to scTRAIL fusion	Increased antitumour activity on colorectal cancer xenografts in mice	211
scFc-Fc-scTRAIL	EGFR	Cytotoxic activity similar to scFv-EHD2- scTRAIL	Superior anticancer activity on colorectal cancer xenografts in mice	212
Fc-scTRAIL	Untagged	Antitumour activity in colorectal cancer cell lines	Strong antitumour activity	213
scFvhu225-Fc- scTRAIL	EGFR	Antitumour activity in colorectal cancer cell lines	High antitumour activity	213
scFv4D5-Fc-scTRAIL	HER2	Antitumour activity in colorectal cancer cell lines	-	213
(scFv3-43-Fc- scTRAIL	HER3 (human epidermal growth factor receptor 3)	Antitumour activity in colorectal cancer cell lines	Strongest antitumour activity	213
scFv323/A3hu3-Fc- scTRAIL	EpCAM (epithelial cell adhesion molecule)	Antitumour activity in colorectal cancer cell lines	Moderate antitumour activity	213
IL2-αCD38-αCD38- scTRAIL	IL2 (interleukin 2)	Selective cell death induction in CD38-positive cancer cells	-	215
VHH-D9-scTRAIL	KV10.1(voltage-gated potassium channel)	Rapid and strong apoptosis induction in different tumour models <i>in vitro</i>	-	216
scFv62-scTRAIL	KV10.1(voltage-gated potassium channel)	Less effectiveness than VHH-D9-scTRAIL	-	216
CD19-TRAIL	CD19 (cluster of differentiation 19)	Pronounced apoptosis induction in CD19- positive BCP-ALL cell lines <i>in vitro</i> and <i>in vivo</i>	Prolonged survival of mice transplanted with BCP-ALL patient-derived xenograft	217

#### Table 3 (continued).

Note. A dash indicates that no data on *in vivo* studies are available.

# 8. Multimeric and bifunctional antibodies targeting TRAIL death receptors

Over the past two decades, a variety of DR4 and DR5 receptor agonistic monoclonal antibodies have been developed. However, clinical trials of single TRAIL DR agonists or their combinations with chemotherapeutic agents have been largely disappointing.<sup>6,218</sup> In murine models *in vivo*, the efficacy of DR5 agonistic antibodies (such as drositumab and conatumumab) appeared to be largely dependent on Fc $\gamma$ R-mediated crosslinking.<sup>219–221</sup> Therefore, novel modified versions of secondgeneration DR agonistic antibodies have been developed, focusing on both DR cross-linking to improve efficacy, and reducing the risk of toxicity due to non-tumour targeting effects (Ref. 21).

Among multimeric antibodies, one of the first was HexaBody-DR5/DR5, an equimolar mixture of two DR5-specific IgG1 antibodies with an Fc-domain mutation that independently enhances antibody hexamerization upon Fc $\gamma$ R-mediated crosslinking. HexaBody-DR5/DR5 has been shown to have potent antitumour activity *in vitro* and *in vivo* in large panels of patient-derived xenograft models of solid cancers.<sup>222</sup> It was also cytotoxic to primary cells derived from bone marrow samples of multiple myeloma patients.<sup>223</sup>

Another example is INBRX-109, a humanized agonistic tetravalent anti-DR5 antibody constructed of two identical camelid-derived sdAbs (heavy chain–only binding domains) linked by a on human immunoglobulin G1 (IgG1)-based Fc

domain. The tetravalent format was claimed to provide a balance between high DR5 clustering efficacy and the avoidance of a potential risk of hepatotoxicity.<sup>224</sup> However, the decavalent agonistic IgM antibody with ten DR5 binding sites, IGM-8444, lacked the hepatotoxicity *in vitro* and was highly cytotoxic in a broad panel of tumour cell lines through DR5 multimerization. IGM-8444 also inhibited tumour growth in several xenograft tumour models both as monotherapy and in combination with the BCL-2 inhibitor ABT-199, and was particularly effective in a gastric PDX model.<sup>67</sup> All three of the above-mentioned



**Figure 6.** Working model of BaCa antibody. (*a*) Healthy tissues are generally non-responsive to agonist DR5 therapy because they express no or very low level of FOLR1, thus DR5 oligomerization and activation is minimal. (*b*) In heterogeneous FOLR1-expressing OvCa cells *in vitro*, FOLR1 acts as an anchoring ligand to recruit BaCa antibody close to the DR5 antigen at the cell surface in an avidity-optimized manner. This induces a high level of DR5 clustering and activation of the apoptotic pathway in both *cis* and *trans* manner selectively in FOLR1+OvCa cells. (*c*) *In vivo*, tumour-associated leukocytes (TAL) express inhibitory FcyRIIB receptor, which is required for the activity of DR5 agonist antibodies. Once engaged *via* FcyRIIB, the BaCa antibody also crosslinks the initial ternary complex (FcyRIIB-BaCa-DR5) *via* a FOLR1 anchor into a high-affinity stable quaternary complex (FOLR1-FcyRIIB-BaCa-DR5), which not only retains the antibody in the tumour tissue but also induces a highly superior TRAIL-R2 activation. Reproduced from Shivange *et at.*<sup>228</sup> with the permission from Elsevier.

multimeric DR5 agonists subsequently entered the clinical trials listed below in Section 9.

Alternatively, to further improve the therapeutic efficacy of anti-DR5 antibodies, several bifunctional antibodies with DR5 specificity of at least one of the moieties have been developed. Two bispecific antibodies, C-BsAb-SS/GS4 and N-BsAb-SS/GS4, were generated by fusing the anti-LT $\beta$ R (lymphotoxin- $\beta$  receptor) single-chain Fv (scFv) to either N- or C-terminus of the heavy chain of the anti-DR5 antibody 14A2. Both variants inhibited tumour growth of LT $\beta$ R-expressing cells *in vivo*.<sup>225</sup>

The MCSPxDR5 bispecific tetravalent antibody was engineered by covalently linking a high affinity MCSP (melanoma-associated chondroitin sulfate proteoglycan) mAb complemented with a human IgG1 Fc domain to the variable binding domains of the DR5 agonistic antibody tigatuzumab for the treatment of melanoma. MCSP (CSPG4) is a promising target for cancer therapy as it is overexpressed in several malignancies and is involved in tumour growth, survival and metastasis.<sup>180</sup> MCSPxDR5 exhibited high affinity for MCSP, resulting in the activation of DR5 in melanoma cells. Antitumour activity of MCSPxDR5 was enhanced by FcγR-mediated crosslinking by myeloid immune effector cells.<sup>181</sup>

The engineered FAP-DR5 tetravalent antibody RG7386 is a bispecific antibody that simultaneously targets both fibroblastactivation protein (FAP) on cancer-associated fibroblasts in the tumour stroma and DR5 on tumour cells. FAP-driven binding of RG7386 mediated the high level of DR5 clustering required to trigger cell death. Antitumour efficacy of RG7386 was strongly FAP-dependent, and independent of Fc $\gamma$ R-crosslinking. Treatment with RG7386 resulted in tumour cell apoptosis *in vitro* and significant tumour regression in several xenograft models, including patient-derived xenograft models with FAP expression either on the stroma or on malignant cells.<sup>226</sup>

Another research group<sup>227</sup> created a humanized scDB, the bispecific antibody targeting DR5 and CD3 with very low aggregate content and high stability and functionality. scDB triggered DR5 activation, and importantly, its anticancer activity was dramatically enhanced by redirecting cytotoxic T cells against both sensitive and resistant melanoma cells.

BaCa (bispecific-anchored cytotoxicity activator) is a singleagent bispecific antibody that simultaneously targets folic acid receptor alpha 1 (FOLR1) and DR5 on FOLR1-expressing ovarian cancer cells. Detailed studies showed that BaCa antibodies derived from AMG-655, lexatumumab or antimurine DR5 antibody MD5-1 were 100 times more effective in inducing cytotoxicity *in vitro* than their parental counterparts. The authors hypothesized that the stronger antitumour response *in vitro* or *in vivo* may be due to BaCa supporting FcgRIIB cross-linking (Fig. 6).<sup>228</sup>

Based on the anti-DR5 antibody TAS266 (Ref. 229), an optimized bispecific antibody BI 905711(CDH17:TRAILR2)<sup>230</sup> which binds to CDH17 (cadherin-17 from the cadherin superfamily of adhesion molecules) and DR5 receptor, was created to reduce the drug-induced liver toxicity of this DR

Table 4. Multimeric and bifunctional antibodies targeting DR5 receptors.

Name	Structure	Effects	In vivo activity	Ref.
	Multim	eric antibodies to DR5		
HexaBody-DR5/DR5	Equimolar mixture of two IgG1 antibodies binding non-overlapping DR5 epitopes with Fc-domain mutation for antibody hexamerization	Potent antitumour activity <i>in</i> <i>vitro</i> and <i>in vivo</i> in large panels xenograft models of solid cancers	Xenograft models of melanoma non-small cells lung cancer, (NSCLC), colorectal, pancreatic, gastric cancers	,223
INBRX-109	Humanized agonistic tetravalent anti-DR5 antibody built of two identical camelid-derived sdAbs joined with an Fc domain of human IgG1	Antitumour activity <i>in vitro</i> in chondrosarcoma cell line and <i>in vivo</i> in chondrosarcoma PDX models	Patient-derived chondro- sarcoma xenograft tumours	224
IGM-8444	Decavalent agonistic anti-DR5 IgM antibody	High cytotoxicity in a broad panel of solid and hematologic cancer cell lines without killing primary human hepatocytes <i>in vitro</i> . Inhibition of tumour growth in multiple xenograft tumour models	Xenograft models of NSCLC, colorectal and gastric cancers	67
	Bifunct	ional antibodies to DR5		
C-BsAb-SS/GS4 N-BsAb-SS/GS4	DR5 and LTβR (Lymphotoxin-beta Receptor)	Inhibition of LTβR-positive tumour growth <i>in vivo</i>	Xenografts models of colon and breast carcinomas	225
MCSPxDR5	DR5 and MCSP (chondroitin sulfate proteoglycan 4)	High cytotoxicity in melanoma cell lines	-	181
RG7386, FAP-DR5	DR5 and fibroblast-activation protein (FAP)	Potent tumour cell apoptosis <i>in vitro</i> and <i>in vivo</i>	Colorectal and breast cancer xenografts model, FAP-positive mesenchymal tumours	226
scDB	DR5 and CD3 (cluster of differentiation 3)	Dramatical potentiation of anticancer activity by the redirection of cytotoxic T cells	-	227
BaCa-1	DR5 and FOLR1 (folate receptor alpha-1)	>100-fold higher <i>in vitro</i> cytotoxicity than parental counterparts	Ovarian cancer xenograft model	228
BI 905711	DR5 and CDH17 (cadherin superfamily of adhesion molecules)	Selective antitumour activity in CDH17-positive colorectal cancer cells	CDH17-positive colorectal cancer xenograft mouse models	230

Note. A dash indicates that no data on in vivo studies are available.

agonist. The membrane cell adhesion protein CDH17 is predominantly expressed in intestinal epithelial cells and is thought to regulate the direction and efficiency of epithelial water transport through trans-interactions with cadherins of neighboring cells.<sup>231</sup> It was found that BI 905711 potently triggered the extrinsic apoptosis pathway and was highly effective in several CDH17-positive colorectal xenograft models.<sup>230</sup> The clinical trials of BI 905711 as well as the aforementioned FAP-DR5 tetravalent antibody RG7386 are discussed below in Section 9. A summary of multimeric and

bifunctional antibodies targeting DR5 receptors is presented in Table 4.

### 9. Clinical trials of next generation TRAIL death receptor agonists

Among the numerous newly developed next-generation TRAIL death receptor agonists, only a few have completed or are in clinical trials (Table 5). In November 2023, Aponermin (circularly permuted TRAIL, CPT) developed by Sunbio

Table 5. Summary of clinical trials of DR-targeting fusions and multimeric preparations.

Name, structure	Study title	Setting	Stage	Clinical trial identifier	
Aponermin (Circularly Permuted	A Phase Ib dose escalation study of recombinant circularly permuted TRAIL in patients with relapsed or refractory multiple myeloma	Monotherapy	Phase Ib	ChiCTR-TNRC-12001896	
TRAIL, CPT)	Phase II open-label study of recombinant circularly permuted TRAIL as a single-agent treatment for relapsed or refractory multiple myeloma	Monotherapy	Phase II	ChiCTR-ONC-12002065	
	A multicenter, open-label Phase II study of recombinant CPT (circularly permuted TRAIL) plus thalidomide in patients with relapsed and refractory multiple myeloma	Thalidomide	Phase II	ChiCTR-ONC-12002066	
	Circularly permuted TRAIL plus thalidomide and dexamethasone versus thalidomide and dexamethasone for relapsed/refractory multiple myeloma: a Phase II study	Thalidomide Dexamethasone	Phase II	ChiCTR-TRC-11001625	
	A multicenter, randomized, double-blind, controlled Phase III study of CPT or placebo in combination with thalidomide and dexamethasone in subjects with relapsed or refractory multiple myeloma	Thalidomide Dexamethasone	Phase III	ChiCTR-IPR-15006024	
Rilunermin alfa (SCB-313)	Study with SCB-313 (recombinant human TRAIL-trimer fusion protein) for treatment of peritoneal malignancies	Monotherapy	Phase I	NCT03443674	
	Study with SCB-313 (recombinant human trail-trimer fusion protein) for treatment of malignant ascites	Monotherapy	Phase I	NCT04051112	
	A Phase I study evaluating SCB-313 (recombinant human trail-trimer fusion protein) for the treatment of malignant pleural effusion	Monotherapy	Phase I	NCT04123886	
	Study with SCB-313 (recombinant human TRAIL-trimer fusion protein) for treatment of malignant pleural effusions	Monotherapy	Phase I	NCT03869697	
	A Phase I study evaluating SCB-313 for the treatment of subjects with peritoneal carcinomatosis	Monotherapy	Phase I	NCT04047771	
Eftozanermin alfa (ABBV-621)	A study of the safety and tolerability of ABBV-621 in participants with previously-treated solid tumours and hematologic malignancies	Venetoclax Bevacizumab FOLFIRI	Phase I	NCT03082209	
	Eftozanermin alfa in combination with bortezomib and dexamethasone in adult participants with relapsed or refractory multiple myeloma	Bortezomib Dexamethasone	Phase II	NCT04570631	
RO6874813 (RG7386, FAP-DR5)	A dose-escalation study of RO6874813 in participants with locally advanced or metastatic solid tumours	Monotherapy	Phase I	NCT02558140	
BI 905711	A study to find a safe and effective dose of BI 905711 in patients with advanced gastrointestinal cancer	Monotherapy	Phase Ia/b	NCT04137289	
Benufutamab – Genmab (GEN1029, HexaBody-DR5/DR5	First-in-human, open-label, dose-escalation trial with expansion cohorts to evaluate safety of GEN1029 in patients with malignant solid tumours	Monotherapy	Phase I/II	NCT03576131	
Ozekibart (INBRX-109)	An open-label, multicenter, first-in-human, phase 1 dose- escalation and multicohort expansion study of INBRX-109 in subjects with locally advanced or metastatic solid tumours including sarcomas	Monotherapy	Phase I	NCT03715933	
	A randomized, blinded, placebo-controlled, Phase II study of INBRX-109 in unresectable or metastatic conventional chondrosarcoma	Monotherapy	Phase II	NCT04950075	
Aplitabart (IGM-8444)	Phase Ia/Ib clinical trial of Aplitabart as a single agent and in combination in patients with relapsed and/or refractory solid or hematologic cancers	FOLFIRI+ bevacizumab	Phase I	NCT04553692	
Note. CPT is circularly permuted TRAIL; SCB-313 is Recombinant Human TRAIL-Trimer Fusion Protein.					

Biotech Co. Ltd. (China) received its first approval in combination with thalidomide and dexamethasone for the treatment of patients with relapsed or refractory multiple myeloma who have received at least two prior therapies.<sup>232</sup> This can be considered a milestone as Aponermin is the first therapy targeting TRAIL death receptors to receive clinical approval, thereby validating them as a clinically relevant target.

Another stabilized TRAIL variant SCB-313 (drug name Rilunermin alfa) developed by Clover Biopharmaceuticals, Ltd. (Shanghai, China) for the treatment of intracavitary malignancies including malignant ascites, malignant pleural effusions and peritoneal carcinomatosis, has completed Phase I clinical trials.<sup>233</sup>

ABBV-621, Eftozanermin alfa, is a hexameric TRAIL death receptor agonist being developed by AbbVie Inc. (North Chicago, Illinois USA) for the treatment of hematological malignancies and multiple solid tumours in combination with such chemotherapeutic agents as bortezomib and dexamethasone.116 In Phase I trials, Eftosanermin alfa demonstrated acceptable tolerability and safety at a dose of 7.5 mg kg<sup>-1</sup> once weekly, while also providing increased tumour regression in colorectal cancer.234

The bispecific antibody RO6874813 (RG7386, FAP-DR5)<sup>226</sup> has been clinically developed by the pharmaceutical company F. Hoffmann-La Roche AG (Basel, Switzerland). In a Phase I dose-escalation study, RO6874813 demonstrated a favourable safety profile in patients with various types of solid tumours, and preliminary antitumour activity was observed in patients with heavily treated NSCLC.<sup>235</sup>

Recently, the pharmaceutical company Boehringer Ingelheim (Ingelheim am Rhein, Germany) developed and reported the results of an open-label dose-escalation Phase Ia/b study of the bispecific antibody BI 905711 (CDH17:TRAILR2) targeting the cadherin superfamily of adhesion molecules CDH17 and the DR5 receptor (Ref. 230) in patients with advanced gastrointestinal cancer. In pre-treated patients, BI 905711 was associated with an acceptable safety profile and early signs of disease control. It is planned to continue Phase Ib clinical trials with BI 905711 using different dosing regimens (0.6/1.2/2.4 mg kg<sup>-1</sup> every 14 days, and 0.6 mg kg<sup>-1</sup> weekly).<sup>236</sup>

Among the multimeric DR5 agonistic antibodies, GEN1029 (HexaBody-DR5/DR5), named Benufutamab, was launched in 2018 in a Phase I/II clinical trial (NCT03576131) in a mixed population of patients with certain solid tumours, with support from the biotech company Genmab. However, the trial was stopped due to a narrow therapeutic window after the dose-escalation part, as explained by the sponsor.

INBRX-109 (Ozekibart) developed by Inhibrx Biosciences, Inc. (La Jolla, CA, USA) showed significant antitumour activity and a good safety profile in a Phase I trial in patients with unresectable/metastatic chondrosarcoma.<sup>224</sup> A randomized, placebo-controlled, Phase II trial in chondrosarcoma is currently ongoing. Importantly, the trial has received orphan drug designation from both the FDA and EMA (European Medicines Agency).

IGM Biosciences, Inc. (Mountain View, CA, USA) is currently enrolling patients in a Phase Ia/Ib clinical trial of IGM-8444 (Aplitabart)<sup>67</sup> as a single agent and in combination in patients with relapsed and/or refractory solid or hematologic cancers, including colorectal cancer, sarcoma, non-Hodgkin's lymphoma, AML and chronic lymphocytic leukemia.

Finally, a PEGylated recombinant human TRAIL TLY012,<sup>91–93</sup> a lead product candidate from Theraly Fibrosis, Inc. (Baltimore, MD, USA), a subsidiary of D&D Pharmatech

(Seongnam, South Corea), stands apart from cancer treatment. TLY012 has been granted an Orphan Drug Designation for the treatment of chronic pancreatitis in 2019 and systemic sclerosis in 2020. TLY012 is also being investigated for the treatment of non-alcoholic steatohepatitis (NASH) liver fibrosis. This confirms the potential for expanding the spectrum of application of TRAIL receptor agonists to be used not only in the treatment of cancer, but also in other diseases.

### **10.** Conclusion

Clinical evaluation of the first-generation-TRAIL death receptor agonists has failed to demonstrate sufficient efficacy. However, the development of the novel modified versions of nextgeneration TRAIL death receptor agonists with enhanced antitumour properties provides confidence that targeting these receptors will eventually be brought to clinical application. A better understanding of the mechanisms by which resistance to TRAIL-mediated apoptosis signalling develops has led to the creation of new modified proteins or bispecific fusions based on death receptor agonists with potent antitumour properties. Whether next-generation drugs targeting DR4 and DR5 will be approved for cancer therapy depends on the results of preclinical or clinical trials. Particularly promising are bifunctional proteins that simultaneously target tumour cells and other components of the tumour microenvironment (immune cells, the network of tumour vasculature and other non-cellular components) surrounding the tumour are particularly promising, since it is now well established that tumour cell proliferation is largely regulated by components of the microenvironment. Optimization of bispecific protein design, such as diversification of linkerbased adapters and selection of effector molecules, opens up new opportunities to increase the therapeutic index of TRAIL DR agonists.

#### The authors declare no conflicts of interest. Author contributions

Alina A. Isakova, Marine E. Gasparian: writing — original draft;

Artem A. Artykov: visualization;

Dmitry A. Dolgikh: project administration, funding acquisition;

Mikhail P. Kirpichnikov: supervision;

Anne V. Yagolovich: conceptualization, data curation, writing — review and editing.

The study was financially supported by the Russian Science Foundation (Grant No. 24-14-00250, https://rscf.ru/project/24-14-00250/).

### 11. List of abbreviations

ABD — albumin-binding domain;

ADI — arginine deiminase;

Akt — protein kinase B;

AML — acute myeloid leukemia;

AMPs — antimicrobial peptides;

ANG2 — angiopep-2;

APAF1 — apoptotic protease activating factor 1;

BaCa — bispecific-anchored cytotoxicity activator;

Bak — BCL2 antagonist/killer 1;

Bax — member of the BCL-2 gene family;

BBB — blood-brain barrier;

BCL-2 — member of the BCL-2 family of regulator proteins;

BCL-XL — B-cell lymphoma-extra large;

NF-Kb - nuclear factor kappa-light-chain-enhancer of BCP-ALL — B-cell precursor acute lymphoblastic leukemia; Bfl1/A1 — BCL-2-related protein A1; activated B cells: BFPs — bifunctional proteins; NRP-1 — neuropilin-1; BH3 — domain BCL-2 homology domain; NSCLC --- non-small-cell lung cancer; B-NHL — B-cell non-Hodgkin's lymphoma; OPG — osteoprotegerin; BPL — B-cell precursor acute lymphoblastic leukemia; Par-4 — prostate apoptosis response-4; CD19B — lymphocyte antigen; PDGFR $\beta$  — platelet-derived growth factor receptor beta; CD20 — B-lymphocyte antigen; PDL1 — programmed death-ligand 1; CD3 — cluster of differentiation 3; PDXs — patient-derived xenografts; CD33 — myeloid cell surface antigen; PEG — polyethylene glycol; CD40 — TNF receptor superfamily member 5; PI3K — phosphoinositide 3-kinases; CD47 — cluster of differentiation 47; RBC — red blood cell; CD7 — cell surface glycoprotein; RIPK1 — receptor-interacting serine/threonine-protein CD70 — cluster of differentiation 70; kinase 1; RTX — rituximab; CDK9 — cyclin-dependent kinase 9; c-FLIP — FLICE-like inhibitory protein; SAC — selective for apoptosis of cancer cells; cIAP — cellular inhibitor of apoptosis protein-1; scFv — single-chain variable fragment; CLL1 — C-type lectin-like molecule-1; scTRAIL — single-chain TRAIL; CML — chronic myeloid leukemia; SiRNA — small interfering RNA; CNS — central nervous system; Smac — second mitochondria-derived activator of caspase; CPT — circularly permuted TRAIL; Src — family of non-receptor tyrosine kinases; CRC — colorectal carcinoma; SRH—antiangiogenic synthetic peptide SRHTKQRHTALH; CSPG4 — chondroitin sulfate proteoglycan 4; STAT3 — signal transducer and activator of transcription 3; DcR — decoy receptor 1; TGF3L — the third disulfide loop of TGF-A; DD — death domain; TMTP1 — the synthetic 5-amino acid peptide NVVRQ; DIABLO — direct inhibitor of apoptosis-binding protein TNC — chicken tenascin-C; with LOw pI; TNF — vascular endothelial growth factor receptor; DISC — death-inducing signalling complex; TRADD — TNF receptor-associated death domain; DLBCL — diffuse large B-cell lymphoma; TRAF2 — TNF receptor-associated factor 2; DR — death receptor; TRAIL - tumour necrosis factor-related apoptosis-inducing DRD2 — dopamine D2 receptor; ligand: DSG3 — desmoglein-3; TWEAK — TNF-like weak inducer of apoptosis; EGFR — epidermal growth factor receptor; VAS — vasostatin; EGP2 — epithelial glycoprotein 2; VEGFR — vascular endothelial growth factor receptor; EHD2 — EH-domain-containing protein 2; VH - variable heavy chain; ELPs — elastin-like polypeptides; VL - variable light chain; EMA — European medicines agency; XIAP — X-linked inhibitor of apoptosis protein. EpCAM — epithelial cell adhesion molecule; ERK — extracellular signal-regulated kinases; **12. References** FA1FT — avian Ad1 spineless fibre; 1. Y.Du, J.Xu. Adv. Mater., 33 (48), 2103114 (2021); FADD — Fas-associated death domain; https://doi.org/10.1002/adma.202103114 FAP — fibroblast activation protein; 2. X.Zheng, Y.Wu, J.Bi, Y.Huang, Y.Cheng, Y.Li, Y.Wu, G.Cao, FDA — US food and drug administration; Z.Tian. Cell Mol. Immunol., 19 (2), 192 (2022); FOLR1 — folic acid receptor alpha 1; https://doi.org/10.1038/s41423-021-00786-6 GBM — glioblastoma multiforme; 3. A.Ashkenazi, R.C.Pai, S.Fong, S.Leung, D.A.Lawrence, HA5FT — human Ad5 fibre shaft; S.A.Marsters, C.Blackie, L.Chang, A.E.McMurtrey, A.Hebert, hCtr1 — human copper transporter 1; L.DeForge, I.L.Koumenis, D.Lewis, L.Harris, J.Bussiere, HER — human epidermal growth factor receptor; H.Koeppen, Z.Shahrokh, R.H.Schwall. J. Clin. Invest., 104 (2), IAPs — inhibitor of apoptosis proteins; 155 (1999); https://doi.org/10.1172/JCI6926 IC<sub>50</sub> — half-maximal inhibitory concentration; 4. A.Ashkenazi. Nat. Rev. Cancer, 2 (6), 420 (2002); IDH — isocitrate dehydrogenase; https://doi.org/10.1038/nrc821 5. R.S.Herbst, S.G.Eckhardt, R.Kurzrock, S.Ebbinghaus, IL2 — interleukin 2; P.J.O'Dwyer, M.S.Gordon, W.Novotny, M.A.Goldwasser, iRGD — cyclic Peptide CRGDKGPDC; T.M.Tohnya, B.L.Lum, A.Ashkenazi, A.M.Jubb, JAK2 — Janus kinase 2; D.S.Mendelson. J. Clin. Oncol., 28 (17), 2839 (2010); KV10.1 — voltage-gated potassium channel; https://doi.org/10.1200/JCO.2009.25.1991 LDM — Lidamycin; 6. M.Kundu, Y.E.Greer, J.L.Dine, S.Lipkowitz. Cells, 11 (23), LRP1 — low density lipoprotein receptor-related protein 1; 3717 (2022); https://doi.org/10.3390/cells11233717 MAPK — mitogen-activated protein kinase; 7. D.Deng, K.Shah. Trends Cancer, 6 (12), 989 (2020); MCL1 — induced myeloid leukemia cell differentiation https://doi.org/10.1016/j.trecan.2020.06.006 protein; J.-C.Soria, E.Smit, D.Khayat, B.Besse, X.Yang, C.-P.Hsu, D.Reese, J.Wiezorek, F.Blackhall. J. Clin. Oncol., 28 (9), 1527 MCSP — melanoma chondroitin sulfate proteoglycan; (2010); https://doi.org/10.1200/JCO.2009.25.4847 MMAE — monomethyl auristatin E; 9. C.Y.Cheah, D.Belada, M.A.Fanale, A.Janikova,

M.S.Czucman, I.W.Flinn, A.V.Kapp, A.Ashkenazi, S.Kelley,

- MOMP mitochondrial outer membrane permeabilization;
- MRP3 multidrug resistance protein 3;

G.L.Bray, S.Holden, J.F.Seymour. *Lancet Haematol.*, **2** (4), e166 (2015); https://doi.org/10.1016/S2352-3026(15)00026-5

- J.-C.Soria, Z.Márk, P.Zatloukal, B.Szima, I.Albert, É.Juhász, J.-L.Pujol, J.Kozielski, N.Baker, D.Smethurst, Y.Hei, A.Ashkenazi, H.Stern, L.Amler, Y.Pan, F.Blackhall. J. Clin. Oncol., 29 (33), 4442 (2011); https://doi.org/10.1200/JCO.2011.37.2623
- Z.A.Wainberg, W.A.Messersmith, P.F.Peddi, A.V.Kapp, A.Ashkenazi, S.Royer-Joo, C.C.Portera, M.F.Kozloff. *Clin. Colorectal Cancer*, **12** (4), 248 (2013); https://doi.org/10.1016/j.clcc.2013.06.002
- G.P.Amarante-Mendes, T.S.Griffith. *Pharmacol. Ther.*, **155**, 117 (2015); https://doi.org/10.1016/j.pharmthera.2015.09.001
- T.Hartwig, A.Montinaro, S.Von Karstedt, A.Sevko, S.Surinova, A.Chakravarthy, L.Taraborrelli, P.Draber, E.Lafont, F.Arce Vargas, M.A.El-Bahrawy, S.A.Quezada, H.Walczak. *Mol. Cell*, 65 (4), 730 (2017); https://doi.org/10.1016/j.molcel.2017.01.021
- 14. Y.-T.Oh, S.-Y.Sun. *Biomolecules*, **11** (4), 499 (2021); https://doi.org/10.3390/biom11040499
- S.von Karstedt, A.Conti, M.Nobis, A.Montinaro, T.Hartwig, J.Lemke, K.Legler, F.Annewanter, A.D.Campbell, L.Taraborrelli, A.Grosse-Wilde, J.F.Coy, M.A.El-Bahrawy, F.Bergmann, R.Koschny, J.Werner, T.M.Ganten, T.Schweiger, K.Hoetzenecker, I.Kenessey, B.Hegedüs, M.Bergmann, C.Hauser, J.-H.Egberts, T.Becker, C.Röcken, H.Kalthoff, A.Trauzold, K.I.Anderson, O.J.Sansom, H.Walczak. *Cancer Cell*, **27** (4), 561 (2015); https://doi.org/10.1016/j.ccell.2015.02.014
- H.Fritsche, T.Heilmann, R.J.Tower, C.Hauser, A.Von Au, D.El-Sheikh, G.M.Campbell, G.Alp, D.Schewe, S.Hübner, S.Tiwari, D.Kownatzki, S.Boretius, D.Adam, W.Jonat, T.Becker, C.C.Glüer, M.Zöller, H.Kalthoff, C.Schem, A.Trauzold. *Oncotarget.*, 6 (11), 9502 (2015); https://doi.org/10.18632/oncotarget.3321
- K.Azijli, B.Weyhenmeyer, G.J.Peters, S.De Jong, F.A.E.Kruyt. *Cell Death Differ.*, **20** (7), 858 (2013); https://doi.org/10.1038/cdd.2013.28
- A.Guerrache, O.Micheau. *Cells*, **13** (6), 521 (2024); https://doi.org/10.3390/cells13060521
- A.Dubuisson, O.Micheau. Antibodies, 6 (4), 16 (2017); https://doi.org/10.3390/antib6040016
- D.De Miguel, J.Lemke, A.Anel, H.Walczak, L.Martinez-Lostao. *Cell Death Differ.*, 23 (5), 733 (2016); https://doi.org/10.1038/cdd.2015.174
- F.Di Cristofano, A.George, V.Tajiknia, M.Ghandali, L.Wu, Y.Zhang, P.Srinivasan, J.Strandberg, M.Hahn, A.Sanchez Sevilla Uruchurtu, A.A.Seyhan, B.A.Carneiro, L.Zhou, K.E.Huntington, W.S.El-Deiry. *Biochem. Soc. Transactions*, 51 (1), 57 (2023); https://doi.org/10.1042/BST20220098
- A.Montinaro, H.Walczak. Cell Death Differ., 30 (2), 237 (2023); https://doi.org/10.1038/s41418-022-01059-z
- H.Alizadeh Zeinabad, E.Szegezdi. Cancers, 14 (20), 5125 (2022); https://doi.org/10.3390/cancers14205125
- A.V.Yagolovich, M.E.Gasparian, D.A.Dolgikh. *Pharmaceutics*, **15** (2), 515 (2023); https://doi.org/10.3390/pharmaceutics15020515
- S.R.Wiley, K.Schooley, P.J.Smolak, W.S.Din, C.-P.Huang, J.K.Nicholl, G.R.Sutherland, T.D.Smith, C.Rauch, C.A.Smith, R.G.Goodwin. *Immunity*, 3 (6), 673 (1995); https://doi.org/10.1016/1074-7613(95)90057-8
- R.M.Pitti, S.A.Marsters, S.Ruppert, C.J.Donahue, A.Moore, A.Ashkenazi. J. Biol. Chem., 271 (22), 12687 (1996); https://doi.org/10.1074/jbc.271.22.12687
- S.-S.Cha, M.-S.Kim, Y.H.Choi, B.-J.Sung, N.K.Shin, H.-C.Shin, Y.C.Sung, B.-H.Oh. *Immunity*, **11** (2), 253 (1999); https://doi.org/10.1016/S1074-7613(00)80100-4
- J.-L.Bodmer, P.Meier, J.Tschopp, P.Schneider. J. Biol. Chem., 275 (27), 20632 (2000); https://doi.org/10.1074/jbc.M909721199

- S.G.Hymowitz, M.P.O'Connell, M.H.Ultsch, A.Hurst, K.Totpal, A.Ashkenazi, A.M.De Vos, R.F.Kelley. *Biochemistry*, **39** (4), 633 (2000); https://doi.org/10.1021/bi9922421
- N.-B.Liabakk, A.Sundan, S.Torp, P.Aukrust, S.S.Frøland, T.Espevik. J. Immunol. Methods, 259 (1-2), 119 (2002); https://doi.org/10.1016/S0022-1759(01)00501-4
- S.M.Mariani, B.Matiba, E.A.Armandola, P.H.Krammer. J. Cell Biol., 137 (1), 221 (1997); https://doi.org/10.1083/jcb.137.1.221
- P.A.Holoch, T.S.Griffith. *Eur. J. Pharmacol.*, **625** (1–3), 63 (2009); https://doi.org/10.1016/j.ejphar.2009.06.066
- M.E.Gasparian, B.V.Chernyak, D.A.Dolgikh, A.V.Yagolovich, E.N.Popova, A.M.Sycheva, S.A.Moshkovskii, M.P.Kirpichnikov. *Apoptosis*, 14 (6), 778 (2009); https://doi.org/10.1007/s10495-009-0349-3
- H.Walczak. *EMBO J.*, **16** (17), 5386 (1997); https://doi.org/10.1093/emboj/16.17.5386
- G.Pan, K.O'Rourke, A.M.Chinnaiyan, R.Gentz, R.Ebner, J.Ni, V.M.Dixit. *Science*, **276** (5309), 111 (1997); https://doi.org/10.1126/science.276.5309.111
- J.P.Sheridan, S.A.Marsters, R.M.Pitti, A.Gurney, M.Skubatch, D.Baldwin, L.Ramakrishnan, C.L.Gray, K.Baker, W.I.Wood, A.D.Goddard, P.Godowski, A.Ashkenazi. *Science*, 277 (5327), 818 (1997); https://doi.org/10.1126/science.277.5327.818
- M.A.Degli-Esposti, W.C.Dougall, P.J.Smolak, J.Y.Waugh, C.A.Smith, R.G.Goodwin. *Immunity*, 7 (6), 813 (1997); https://doi.org/10.1016/S1074-7613(00)80399-4
- G.Pan, J.Ni, Y.-F.Wei, G.Yu, R.Gentz, V.M.Dixit. Science, 277 (5327), 815 (1997); https://doi.org/10.1126/science.277.5327.815
- S.A.Marsters, J.P.Sheridan, R.M.Pitti, A.Huang, M.Skubatch, D.Baldwin, J.Yuan, A.Gurney, A.D.Goddard, P.Godowski, A.Ashkenazi. *Curr. Biol.*, 7 (12), 1003 (1997); https://doi.org/10.1016/S0960-9822(06)00422-2
- D.Mérino, N.Lalaoui, A.Morizot, P.Schneider, E.Solary, O.Micheau. *Mol. Cell. Biol.*, 26 (19), 7046 (2006); https://doi.org/10.1128/MCB.00520-06
- J.G.Emery, P.McDonnell, M.B.Burke, K.C.Deen, S.Lyn, C.Silverman, E.Dul, E.R.Appelbaum, C.Eichman, R.DiPrinzio, R.A.Dodds, I.E.James, M.Rosenberg, J.C.Lee, P.R.Young. *J. Biol. Chem.*, **273** (23), 14363 (1998); https://doi.org/10.1074/jbc.273.23.14363
- M.R.Sprick, M.A.Weigand, E.Rieser, C.T.Rauch, P.Juo, J.Blenis, P.H.Krammer, H.Walczak. *Immunity*, **12** (6), 599 (2000); https://doi.org/10.1016/S1074-7613(00)80211-3
- R.Koschny, H.Walczak, T.M.Ganten. J. Mol. Med., 85 (9), 923 (2007); https://doi.org/10.1007/s00109-007-0194-1
- K.Yaacoub, R.Pedeux, P.Lafite, U.Jarry, S.Aci-Sèche, P.Bonnet, R.Daniellou, T.Guillaudeux. *Curr. Issues Mol. Biol.*, 46 (1), 710 (2024); https://doi.org/10.3390/cimb46010046
- L.M.Humphreys, J.P.Fox, C.A.Higgins, J.Majkut, T.Sessler, K.McLaughlin, C.McCann, J.Z.Roberts, N.T.Crawford, S.S.McDade, C.J.Scott, T.Harrison, D.B.Longley. *EMBO Rep.*, **21** (3), e49254 (2020); https://doi.org/10.15252/embr.201949254
- 46. J.D.Twomey, S.-R.Kim, L.Zhao, W.P.Bozza, B.Zhang. *Drug Resist. Update*, **19**, 13 (2015); https://doi.org/10.1016/j.drup.2015.02.001
- A.A.Artykov, A.V.Yagolovich, D.A.Dolgikh, M.P.Kirpichnikov, D.B.Trushina, M.E.Gasparian. Front. Cell Dev. Biol., 9, 733688 (2021); https://doi.org/10.3389/fcell.2021.733688
- U.Bertsch, C.Röder, H.Kalthoff, A.Trauzold. *Cell Death Dis.*, 5 (8), e1390 (2014); https://doi.org/10.1038/cddis.2014.351
- U.Mert, A.Adawy, E.Scharff, P.Teichmann, A.Willms, V.Haselmann, C.Colmorgen, J.Lemke, S.von Karstedt, J.Fritsch, A.Trauzold. *Cancers*, **11** (8), 1167 (2019); https://doi.org/10.3390/cancers11081167

23 of 27

- Y.Kojima, M.Nakayama, T.Nishina, H.Nakano, M.Koyanagi, K.Takeda, K.Okumura, H.Yagita. *J. Biol. Chem.*, 286 (50), 43383 (2011); https://doi.org/10.1074/jbc.M111.309377
- V.Haselmann, A.Kurz, U.Bertsch, S.Hübner, M.Olempska-Müller, J.Fritsch, R.Häsler, A.Pickl, H.Fritsche, F.Annewanter, C.Engler, B.Fleig, A.Bernt, C.Röder, H.Schmidt, C.Gelhaus, C.Hauser, J.Egberts, C.Heneweer, A.M.Rohde, C.Böger, U.Knippschild, C.Röcken, D.Adam, H.Walczak, S.Schütze, O.Janssen, F.G.Wulczyn, H.Wajant, H.Kalthoff, A.Trauzold. *Gastroenterology*, **146** (1), 278 (2014); https://doi.org/10.1053/j.gastro.2013.10.009
- K.Huang, J.Zhang, K.L.O'Neill, C.B.Gurumurthy, R.M.Quadros, Y.Tu, X.Luo. *J. Biol. Chem.*, **291** (22), 11843 (2016); https://doi.org/10.1074/jbc.M115.711051
- Y.Deng, Y.Lin, X.Wu. Genes Dev., 16 (1), 33 (2002); https://doi.org/10.1101/gad.949602
- Z.Šarif, B.Tolksdorf, H.Fechner, J.Eberle. *Mol. Carcinogenesis*, **59** (11), 1256 (2020); https://doi.org/10.1002/mc.23253
- S.Gunalp, D.G.Helvaci, A.Oner, A.Bursalı, A.Conforte, H.Güner, G.Karakülah, E.Szegezdi, D.Sag. *Front. Immunol.*, 14, 1209249 (2023); https://doi.org/10.3389/fimmu.2023.1209249
- E.Varfolomeev, H.Maecker, D.Sharp, D.Lawrence, M.Renz, D.Vucic, A.Ashkenazi. *J. Biol. Chem.*, 280 (49), 40599 (2005); https://doi.org/10.1074/jbc.M509560200
- A.V.Yagolovich, A.A.Artykov, T.A.Karmakova, M.S.Vorontsova, A.A.Pankratov, A.A.Andreev-Andrievsky, D.A.Dolgikh, M.P.Kirpichnikov, M.E.Gasparian. *Trans. Oncol.*, **13** (4), 100762 (2020); https://doi.org/10.1016/j.tranon.2020.100762
- F.J.H.Hoogwater, M.W.Nijkamp, N.Smakman, E.J.A.Steller, B.L.Emmink, B.F.Westendorp, D.A.E.Raats, M.R.Sprick, U.Schaefer, W.J.Van Houdt, M.T.De Bruijn, R.C.J.Schackmann, P.W.B.Derksen, J.Medema, H.Walczak, I.H.M.Borel Rinkes, O.Kranenburg. *Gastroenterology*, **138** (7), 2357 (2010); https://doi.org/10.1053/j.gastro.2010.02.046
- X.Yuan, A.Gajan, Q.Chu, H.Xiong, K.Wu, G.S.Wu. *Cancer* Metastasis Rev., **37** (4), 733 (2018); https://doi.org/10.1007/s10555-018-9728-y
- Q.Deng, L.Chen, G.Zhang, L.Liu, S.-M.Luo, X.Gao. Int. Immunopharmacol., 138, 112570 (2024); https://doi.org/10.1016/j.intimp.2024.112570
- A.A.Artykov, D.A.Belov, V.O.Shipunova, D.B.Trushina, S.M.Deyev, D.A.Dolgikh, M.P.Kirpichnikov, M.E.Gasparian. *Cancers*, 12 (5), 1129 (2020); https://doi.org/10.3390/cancers12051129
- C.T.Hellwig, M.E.Delgado, J.Skoko, L.Dyck, C.Hanna,
   A.Wentges, C.Langlais, C.Hagenlocher, A.Mack, D.Dinsdale,
   K.Cain, M.MacFarlane, M.Rehm. *Cell Death Differ.*, 29 (1), 147 (2022); https://doi.org/10.1038/s41418-021-00843-7
- L.De Wilt, B.K.Sobocki, G.Jansen, H.Tabeian, S.De Jong, G.J.Peters, F.Kruyt. *Cancer Drug Resist.*, (2024); https://doi.org/10.20517/cdr.2024.14
- 64. J.Montero, R.Haq. *Cancer Discov.*, **12** (5), 1217 (2022); https://doi.org/10.1158/2159-8290.CD-21-1334
- J.H.Song, K.Kandasamy, A.S.Kraft. J. Biol. Chem., 283 (36), 25003 (2008); https://doi.org/10.1074/jbc.M802511200
- 66. G.Wang, Y.Zhan, H.Wang, W.Li. *Cancer Chemother*. *Pharmacol.*, **69** (3), 799 (2012); https://doi.org/10.1007/s00280-011-1763-0
- 67. B.T.Wang, T.Kothambawala, L.Wang, T.J.Matthew, S.E.Calhoun, A.K.Saini, M.F.Kotturi, G.Hernandez, E.W.Humke, M.S.Peterson, A.M.Sinclair, B.A.Keyt. *Mol. Cancer Ther.*, **20** (12), 2483 (2021); https://doi.org/10.1158/1535-7163.MCT-20-1132
- C.Mo, N.Wei, T.Li, M.Ahmed Bhat, M.Mohammadi, C.Kuang. *Biochem. Pharmacol.*, **229**, 116470 (2024); https://doi.org/10.1016/j.bcp.2024.116470

- J.-P.Ruff, A.-L.Kretz, M.Kornmann, D.Henne-Bruns, J.Lemke, B.Traub. *Anticancer Res.*, 41 (12), 5973 (2021); https://doi.org/10.21873/anticanres.15416
- A.Montinaro, I.Areso Zubiaur, J.Saggau, A.-L.Kretz, R.M.M.Ferreira, O.Hassan, E.Kitzig, I.Müller, M.A.El-Bahrawy, S.Von Karstedt, D.Kulms, G.Liccardi, J.Lemke, H.Walczak. *Cell Death Differ.*, 29 (3), 492 (2022); https://doi.org/10.1038/s41418-021-00869-x
- X.Shen, A.-L.Kretz, S.Schneider, U.Knippschild, D.Henne-Bruns, M.Kornmann, J.Lemke, B.Traub. *Biomedicines*, **11** (3), 928 (2023); https://doi.org/10.3390/biomedicines11030928
- V.V.Prabhu, S.Morrow, A.Rahman Kawakibi, L.Zhou, M.Ralff, J.Ray, A.Jhaveri, I.Ferrarini, Y.Lee, C.Parker, Y.Zhang, R.Borsuk, W.-I.Chang, J.N.Honeyman, F.Tavora, B.Carneiro, A.Raufi, K.Huntington, L.Carlsen, A.Louie, H.Safran, A.A.Seyhan, R.S.Tarapore, L.Schalop, M.Stogniew, J.E.Allen, W.Oster, W.S.El-Deiry. *Neoplasia*, **22** (12), 725 (2020); https://doi.org/10.1016/j.neo.2020.09.005
- I.Arrillaga-Romany, S.L.Gardner, Y.Odia, D.Aguilera, J.E.Allen, T.Batchelor, N.Butowski, C.Chen, T.Cloughesy, A.Cluster, J.De Groot, K.S.Dixit, J.J.Graber, A.M.Haggiagi, R.A.Harrison, A.Kheradpour, L.B.Kilburn, S.C.Kurz, G.Lu, T.J.MacDonald, M.Mehta, A.S.Melemed, P.L.Nghiemphu, S.C.Ramage, N.Shonka, A.Sumrall, R.S.Tarapore, L.Taylor, Y.Umemura, P.Y.Wen. J. Clin. Oncol., 42 (13), 1542 (2024); https://doi.org/10.1200/JCO.23.01134
- J.Wagner, C.L.Kline, M.D.Ralff, A.Lev, A.Lulla, L.Zhou, G.L.Olson, B.R.Nallaganchu, C.H.Benes, J.E.Allen, V.V.Prabhu, M.Stogniew, W.Oster, W.S.El-Deiry. *Cell Cycle*, **16** (19), 1790 (2017); https://doi.org/10.1080/15384101.2017.1325046
- S.El-Soussi, R.Hanna, H.Semaan, A.-R.Khater, J.Abdallah, W.Abou-Kheir, T.Abou-Antoun. *Front. Pediatr.*, 9, 693145 (2021); https://doi.org/10.3389/fped.2021.693145
- L.Zhou, L.Zhang, J.Zhang, L.J.Wu, S.Zhang, A.George, M.Hahn, H.P.Safran, C.C.Chen, A.A.Seyhan, E.T.Wong, W.S.El-Deiry. Combination of Imipridone ONC201 or ONC206 with Temozolomide and Radiotherapy in Triple ITR Therapy Reduces Intracranial Tumor Burden and Prolongs Survival in an Orthotopic Wild-Type IDH GBM Mouse Model// September 27, 2024;
- https://doi.org/10.1101/2024.09.25.610187 77. E.Morrish, G.Brumatti, J.Silke. *Cells*, **9** (2), 406 (2020); https://doi.org/10.3390/cells9020406
- M.Wu, G.Wang, Z.Zhao, Y.Liang, H.Wang, M.Wu, P.Min, L.Chen, Q.Feng, J.Bei, Y.Zeng, D.Yang. *Mol. Cancer Ther.*, 12 (9), 1728 (2013); https://doi.org/10.1158/1535-7163.MCT-13-0017
- D.Finlay, M.Vamos, M.González-López, R.J.Ardecky,
   S.R.Ganji, H.Yuan, Y.Su, T.R.Cooley, C.T.Hauser, K.Welsh,
   J.C.Reed, N.D.P.Cosford, K.Vuori. *Mol. Cancer Ther.*, 13 (1),
   5 (2014); https://doi.org/10.1158/1535-7163.MCT-13-0153
- E.J.Park, H.D.Kim, E.K.Choi, K.-L.Hoe, D.-U.Kim. *Biochem. Biophys. Res. Commun.*, **533** (3), 289 (2020); https://doi.org/10.1016/j.bbrc.2020.09.031
- 81. B.Thapa, R.Kc, H.Uludağ. *J. Controlled Release*, **326**, 335 (2020); https://doi.org/10.1016/j.jconrel.2020.07.013
- S.-H.Kim, K.Kim, J.G.Kwagh, D.T.Dicker, M.Herlyn, A.K.Rustgi, Y.Chen, W.S.El-Deiry. J. Biol. Chem., 279 (38), 40044 (2004); https://doi.org/10.1074/jbc.M404541200
- 83. D.W.Seol, T.R.Billiar. *Cancer Res.*, **60** (12), 3152 (2000)
- J.Naval, D.De Miguel, A.Gallego-Lleyda, A.Anel, L.Martinez-Lostao. *Cancers*, **11** (4), 444 (2019); https://doi.org/10.3390/cancers11040444
- H.Walczak, R.E.Miller, K.Ariail, B.Gliniak, T.S.Griffith, M.Kubin, W.Chin, J.Jones, A.Woodward, T.Le, C.Smith, P.Smolak, R.G.Goodwin, C.T.Rauch, J.C.L.Schuh, D.H.Lynch. *Nat. Med.*, 5 (2), 157 (1999); https://doi.org/10.1038/5517

- Y.S.Youn, M.J.Shin, S.Y.Chae, C.-H.Jin, T.H.Kim, K.C.Lee. Biotechnol. Lett., 29 (5), 713 (2007); https://doi.org/10.1007/s10529-006-9300-7
- D.V.Rozanov, A.Y.Savinov, V.S.Golubkov, O.L.Rozanova, T.I.Postnova, E.A.Sergienko, S.Vasile, A.E.Aleshin, M.F.Rega, M.Pellecchia, A.Y.Strongin. *Mol. Cancer Ther.*, 8 (6), 1515 (2009); https://doi.org/10.1158/1535-7163.MCT-09-0202
- B. D.Rozanov, P.Spellman, A.Savinov, A.Y.Strongin. *PLoS ONE*, **10** (4), e0122980 (2015); https://doi.org/10.1371/journal.pone.0122980
- R.S.Fadeev, A.V.Chekanov, N.V.Dolgikh, V.S.Akatov. Biochem. Moscow Suppl. Ser. A, 7 (1), 29 (2013); https://doi.org/10.1134/S1990747812060049
- V.Gupta, S.Bhavanasi, M.Quadir, K.Singh, G.Ghosh, K.Vasamreddy, A.Ghosh, T.J.Siahaan, S.Banerjee, S.K.Banerjee. J. Cell Commun. Signal., 13 (3), 319 (2019); https://doi.org/10.1007/s12079-018-0492-0
- S.Y.Chae, T.H.Kim, K.Park, C.-H.Jin, S.Son, S.Lee, Y.S.Youn, K.Kim, D.-G.Jo, I.C.Kwon, X.Chen, K.C.Lee. *Mol. Cancer Ther.*, 9 (6), 1719 (2010); https://doi.org/10.1158/1535-7163.MCT-09-1076
- Y.Oh, O.Park, M.Swierczewska, J.P.Hamilton, J.Park, T.H.Kim, S.Lim, H.Eom, D.G.Jo, C.Lee, R.Kechrid, P.Mastorakos, C.Zhang, S.K.Hahn, O.Jeon, Y.Byun, K.Kim, J.Hanes, K.C.Lee, M.G.Pomper, B.Gao, S.Lee. *Hepatology*, 64 (1), 209 (2016); https://doi.org/10.1002/hep.28432
- J.-S.Park, Y.Oh, Y.J.Park, O.Park, H.Yang, S.Slania, L.K.Hummers, A.A.Shah, H.-T.An, J.Jang, M.R.Horton, J.Shin, H.C.Dietz, E.Song, D.H.Na, E.J.Park, K.Kim, K.C.Lee, V.V.Roschke, J.Hanes, M.G.Pomper, S.Lee. *Nat. Commun.*, **10**(1), 1128 (2019); https://doi.org/10.1038/s41467-019-09101-4
- Https://doi.org/10.103/s140/019091014
   L.-Q.Pan, H.-B.Wang, J.Lai, Y.-C.Xu, C.Zhang, S.-Q.Chen. *Biomaterials*, 34 (36), 9115 (2013); https://doi.org/10.1016/j.biomaterials.2013.08.020
- billorg/10.1010/j.0000aternais/2015/00.020
   L.Pan, H.Wang, Z.Xie, Z.Li, X.Tang, Y.Xu, C.Zhang, H.Naranmandura, S.Chen. *Adv. Mater.*, 25 (34), 4718 (2013); https://doi.org/10.1002/adma.201301385
- L.-Q.Pan, W.-B.Zhao, J.Lai, D.Ding, X.-Y.Wei, Y.-Y.Li, W.-H.Liu, X.-Y.Yang, Y.-C.Xu, S.-Q.Chen. *Sci. Rep.*, 5 (1), 14872 (2015); https://doi.org/10.1038/srep14872
- L.Q.Pan, Z.M.Xie, X.J.Tang, M.Wu, F.R.Wang, H.Naranmandura, S.Q.Chen. *Appl. Microbiol. Biotechnol.*, 97 (16), 7253 (2013); https://doi.org/10.1007/s00253-012-4604-0
- H.Liu, D.Su, J.Zhang, S.Ge, Y.Li, F.Wang, M.Gravel, A.Roulston, Q.Song, W.Xu, J.G.Liang, G.Shore, X.Wang, P.Liang. *Sci. Rep.*, 7 (1), 8953 (2017); https://doi.org/10.1038/s41598-017-09518-1
- J.Yan, L.Wang, Z.Wang, Z.Wang, B.Wang, R.Zhu, J.Bi, J.Wu, H.Zhang, H.Wu, B.Yu, W.Kong, X.Yu. *Cell Death Dis.*, 7 (6), e2274 (2016); https://doi.org/10.1038/cddis.2016.177
- R.Li, H.Yang, D.Jia, Q.Nie, H.Cai, Q.Fan, L.Wan, L.Li, X.Lu. J. Control. Release, 228, 96 (2016); https://doi.org/10.1016/j.jconrel.2016.03.004
- 101. H.Yang, Y.Feng, H.Cai, D.Jia, H.Li, Z.Tao, Y.Zhong, Z.Li, Q.Shi, L.Wan, L.Li, X.Lu. *Theranostics*, 8 (9), 2459 (2018); https://doi.org/10.7150/thno.23880
- 102. H.Wang, J.S.Davis, X.Wu. Mol. Cancer Ther., 13 (3), 643 (2014); https://doi.org/10.1158/1535-7163.MCT-13-0645
- E.Brin, K.Wu, E.Dagostino, M.Meng-Chiang Kuo, Y.He, W.-J.Shia, L.-C.Chen, M.Stempniak, R.Hickey, R.Almassy, R.Showalter, J.Thomson. *Oncotarget*, 9 (97), 36914 (2018); https://doi.org/10.18632/oncotarget.26398
- 104. F.Fang, A.Wang, S.Yang. Acta Pharmacol. Sin., 26 (11), 1373 (2005); https://doi.org/10.1111/j.1745-7254.2005.00206.x
- 105. Y.Leng, L.Qiu, J.Hou, Y.Zhao, X.Zhang, S.Yang, H.Xi, Z.Huang, L.Pan, W.Chen. *Chin. J. Cancer*, **35** (1), 86 (2016); https://doi.org/10.1186/s40880-016-0140-0
- 106. Z.Xia, Y.Leng, B.Fang, Y.Liang, W.Li, C.Fu, L.Yang, X.Ke, H.Jiang, J.Weng, L.Liu, Y.Zhao, X.Zhang, Z.Huang, A.Liu,

Q.Shi, Y.Gao, X.Chen, L.Pan, Z.Cai, Z.Wang, Y.Wang, Y.Fan, M.Hou, Y.Ma, J.Hu, J.Liu, J.Zhou, X.Zhang, H.Meng, X.Lu, F.Li, H.Ren, B.Huang, Z.Shao, H.Zhou, Y.Hu, S.Yang, X.Zheng, P.Wei, H.Pang, W.Yu, Y.Liu, S.Gao, L.Yan, Y.Ma, H.Jing, J.Du, W.Ling, J.Zhang, W.Sui, F.Wang, X.Li, W.Chen. *BMC Cancer*, **23** (1), 980 (2023); https://doi.org/10.1186/s12885-023-11489-8

- D.Berg, M.Lehne, N.Müller, D.Siegmund, S.Münkel, W.Sebald, K.Pfizenmaier, H.Wajant. *Cell Death Differ.*, 14 (12), 2021 (2007); https://doi.org/10.1038/sj.cdd.4402213
- D.Berg, T.Stühmer, D.Siegmund, N.Müller, T.Giner,
   O.Dittrich-Breiholz, M.Kracht, R.Bargou, H.Wajant. *FEBS J.*,
   276 (23), 6912 (2009); https://doi.org/10.1111/j.1742-4658.2009.07388.x
- 109. D.Spitzer, J.E.McDunn, S.Plambeck-Suess, P.S.Goedegebuure, R.S.Hotchkiss, W.G.Hawkins. *Mol. Cancer Ther.*, 9 (7), 2142 (2010); https://doi.org/10.1158/1535-7163.MCT-10-0225
- 110. M.Huang, H.Zhu, C.Yi, J.Yan, L.Wei, X.Yang, S.Chen, Y.Huang. *Cancer Chemother: Pharmacol.*, **82** (5), 829 (2018); https://doi.org/10.1007/s00280-018-3658-9
- 111. M.Huang, C.Yi, X.-Z.Huang, J.Yan, L.-J.Wei, W.-J.Tang, S.-C.Chen, Y.Huang. Oncol. Lett., **21** (6), 438 (2021); https://doi.org/10.3892/ol.2021.12699
- 112. C.-H.Lee, K.-J.Park, E.-S.Sung, A.Kim, J.-D.Choi, J.-S.Kim, S.-H.Kim, M.-H.Kwon, Y.-S.Kim. *Proc. Natl. Acad. Sci. USA*, **107** (21), 9567 (2010); https://doi.org/10.1073/pnas.1001541107
- 113. C.-H.Lee, K.-J.Park, S.J.Kim, O.Kwon, K.J.Jeong, A.Kim, Y.-S.Kim. J. Mol. Biol., 411 (1), 201 (2011); https://doi.org/10.1016/j.jmb.2011.05.040
- 114. C.Gieffers, M.Kluge, C.Merz, J.Sykora, M.Thiemann, R.Schaal, C.Fischer, M.Branschädel, B.A.Abhari, P.Hohenberger, S.Fulda, H.Fricke, O.Hill. *Mol. Cancer Ther.*, 12 (12), 2735 (2013); https://doi.org/10.1158/1535-7163.MCT-13-0323
- 115. K.Legler, C.Hauser, J.-H.Egberts, A.Willms, C.Heneweer, S.Boretius, C.Röcken, C.-C.Glüer, T.Becker, M.Kluge, O.Hill, C.Gieffers, H.Fricke, H.Kalthoff, J.Lemke, A.Trauzold. *Cell Death Dis.*, 9 (5), 445 (2018); https://doi.org/10.1038/s41419-018-0478-0
- 116. D.C.Phillips, F.G.Buchanan, D.Cheng, L.R.Solomon, Y.Xiao, J.Xue, S.K.Tahir, M.L.Smith, H.Zhang, D.Widomski, V.C.Abraham, N.Xu, Z.Liu, L.Zhou, E.DiGiammarino, X.Lu, N.Rudra-Ganguly, B.Trela, S.E.Morgan-Lappe. *Cancer Res.*, **81** (12), 3402 (2021); https://doi.org/10.1158/0008-5472.CAN-20-2178
- H.Yang, H.Li, F.Yang, Z.Tao, Q.Shi, T.She, Y.Feng, Z.Li, J.Chen, Y.Zhong, T.Su, W.Zeng, Y.Zhang, S.Wang, L.Li, T.Long, D.Long, J.Cheng, H.Zhu, X.Lu. *Biomaterials*, 295, 121994 (2023);
- https://doi.org/10.1016/j.biomaterials.2023.121994
  118. T.She, F.Yang, S.Chen, H.Yang, Z.Tao, H.Xing, J.Chen, H.Chang, H.Lu, T.Su, Y.Jin, Y.Zhong, J.Cheng, H.Zhu, X.Lu. *J. Control. Release*, **361**, 856 (2023); https://doi.org/10.1016/j.jconrel.2023.07.042
- 119. Y.Du, J.Xu. Adv. Mater., 33 (48), 2103114 (2021); https://doi.org/10.1002/adma.202103114
- 120. S.P.Somasekharan, M.Koc, A.Morizot, O.Micheau,
   P.H.B.Sorensen, O.Gaide, L.Andera, J.-C.Martinou. *Apoptosis*,
   18 (3), 324 (2013); https://doi.org/10.1007/s10495-012-0782-6
- 121. J.S.Desgrosellier, D.A.Cheresh. *Nat. Rev. Cancer*, **10** (1), 9 (2010); https://doi.org/10.1038/nrc2748
- H.Javid, M.A.Oryani, N.Rezagholinejad, A.Esparham, M.Tajaldini, M.Karimi-Shahri. *Cancer Med.*, **13** (2), e6800 (2024); https://doi.org/10.1002/cam4.6800
- 123. L.Cao, P.Du, S.-H.Jiang, G.-H.Jin, Q.-L.Huang, Z.-C.Hua. *Mol. Cancer Ther.*, 7 (4), 851 (2008); https://doi.org/10.1158/1535-7163.MCT-07-0533
- 124. X.Wang, X.Qiao, Y.Shang, S.Zhang, Y.Li, H.He, S.Chen. *Amino Acids*, 49 (5), 931 (2017); https://doi.org/10.1007/s00726-017-2395-4

- 125. K.Huang, N.Duan, C.Zhang, R.Mo, Z.Hua. Sci. Rep., 7 (1), 41904 (2017); https://doi.org/10.1038/srep41904
- 126. L.Yin, X.Li, R.Wang, Y.Zeng, Z.Zeng, T.Xie. Int. J. Pept. Res. Ther., **29** (4), 53 (2023);
- https://doi.org/10.1007/s10989-023-10523-4 127. S.Kang, S.Lee, S.Park. *Polymers*, **12** (9), 1906 (2020); https://doi.org/10.3390/polym12091906
- 128. Y.Huang, X.Li, H.Sha, L.Zhang, X.Bian, X.Han, B.Liu. Sci. Rep., 7 (1), 579 (2017); https://doi.org/10.1038/s41598-017-00688-6
- A.V.Yagolovich, A.A.Isakova, A.A.Artykov, Y.V.Vorontsova, D.V.Mazur, N.V.Antipova, M.S.Pavlyukov, M.I.Shakhparonov, A.M.Gileva, E.A.Markvicheva, E.A.Plotnikova, A.A.Pankratov, M.P.Kirpichnikov, M.E.Gasparian, D.A.Dolgikh. *Int. J. Mol. Sci.*, 23 (20), 12687 (2022); https://doi.org/10.3390/ijms232012687
- Z.Li, T.She, H.Yang, T.Su, Q.Shi, Z.Tao, Y.Feng, F.Yang, J.Cheng, X.Lu. *Drug Delivery*, **29** (1), 1698 (2022); https://doi.org/10.1080/10717544.2022.2079766
- 131. G.Hu, W.Zeng, Y.Xia. *Tumor Biol.*, **39** (6), 101042831771462 (2017); https://doi.org/10.1177/1010428317714624
- M.Razmara, B.Hilliard, A.K.Ziarani, R.Murali, S.Yellayi, M.Ghazanfar, Y.H.Chen, M.L.Tykocinski. *Am. J. Pathol.*, **174** (2), 460 (2009); https://doi.org/10.2353/ajpath.2009.080462
- A.Aronin, S.Amsili, T.B.Prigozhina, K.Tzdaka, J.Rachmilewitz, N.Shani, M.L.Tykocinski, M.Dranitzki Elhalel. *PLoS ONE*, 8 (10), e77050 (2013); https://doi.org/10.1371/journal.pone.0077050
- 134. F.Qiu, M.Hu, B.Tang, X.Liu, H.Zhuang, J.Yang, Z.-C.Hua. *Sci. Rep.*, **3** (1), 3565 (2013); https://doi.org/10.1038/srep03565
- 135. G.Garg, J.Gibbs, B.Belt, M.A.Powell, D.G.Mutch, P.Goedegebuure, L.Collins, D.Piwnica-Worms, W.G.Hawkins, D.Spitzer. *BMC Cancer*, 14 (1), 35 2014); https://doi.org/10.1186/1471-2407-14-35
- 136. Y.Su, K.Tatzel, X.Wang, B.Belt, P.Binder, L.Kuroki, M.A.Powell, D.G.Mutch, W.G.Hawkins, D.Spitzer. *Oncotarget*, 7 (21), 31534 (2016); https://doi.org/10.18632/oncotarget.8925
- 137. W.Yang, D.Luo, S.Wang, R.Wang, R.Chen, Y.Liu, T.Zhu, X.Ma, R.Liu, G.Xu, L.Meng, Y.Lu, J.Zhou, D.Ma. *Clin. Cancer Res.*, **14** (17), 5494 2008); https://doi.org/10.1158/1078-0432.CCR-08-0233
- R.Liu, X.Ma, H.Wang, Y.Xi, M.Qian, W.Yang, D.Luo, L.Fan, X.Xia, J.Zhou, L.Meng, S.Wang, D.Ma, L.Xi. *J. Mol Med.*, **92** (2), 165 (2014); https://doi.org/10.1007/s00109-013-1093-2
- 139. J.S.Pieczykolan, K.Kubiński, M.Masłyk, S.D.Pawlak, A.Pieczykolan, P.K.Rózga, M.Szymanik, M.Gałązka, M.Teska-Kamińska, B.Żerek, K.Bukato, K.Poleszak, A.Jaworski, W.Strożek, R.Świder, R.Zieliński. *Invest. New Drugs*, **32** (6), 1155 (2014); https://doi.org/10.1007/s10637-014-0153-y
- 140. F.M.Uckun, D.E.Myers, S.Qazi, Z.Ozer, R.Rose, O.J.D'Cruz, H.Ma. J. Clin. Invest., 125 (3), 1006 (2015); https://doi.org/10.1172/JCI76610
- 141. Y.Zhen, Y.Lin, Y.Li, Y.Zhen. Acta Pharmacol. Sin., 30 (7), 1025 (2009); https://doi.org/10.1038/aps.2009.75
- 142. D.Zhu, X.Wang, Y.Shang, Y.Li, W.Jiang, L.Li, S.Chen. Anti-Cancer Drugs, 26 (1), 64 (2015); https://doi.org/10.1097/CAD.00000000000160
- 143. T.Jiang, C.Zhou, S.Ren. OncoImmunology, 5 (6), e1163462 (2016); https://doi.org/10.1080/2162402X.2016.1163462
- 144. J.Madhumathi, S.Sridevi, R.S.Verma. *Targ. Oncol.*, **11** (4), 535 (2016); https://doi.org/10.1007/s11523-016-0424-y
- Z.Dong, X.Zhang, Q.Zhang, J.Tangthianchaichana, M.Guo, S.Du, Y.Lu. *IJN*, **19**, 1017 (2024); https://doi.org/10.2147/IJN.S445333
- 146. M.Sang, J.Zhang, B.Li, Y.Chen. Protein Expression and Purification, **122**, 82 (2016); https://doi.org/10.1016/j.pep.2016.02.015

- 147. H.Liu, Y.Han, H.Fu, M.Liu, J.Wu, X.Chen, S.Zhang, Y.Chen. *Appl. Microbiol. Biotechnol.*, **97** (7), 2877 (2013); https://doi.org/10.1007/s00253-012-4541-y
- 148. I.Rady, I.A.Siddiqui, M.Rady, H.Mukhtar. *Cancer Lett.*, 402, 16 (2017); https://doi.org/10.1016/j.canlet.2017.05.010
- A.R.Cheratta, F.Thayyullathil, S.Pallichankandy, K.Subburayan, A.Alakkal, S.Galadari. *Cell Death Dis.*, **12** (1), 47 (2021); https://doi.org/10.1038/s41419-020-03292-1
- J.Zhang, W.Dong, Y.Ren, D.Wei. Appl. Microbiol. Biotechnol., 106 (4), 1511 2022); https://doi.org/10.1007/s00253-022-11807-3
- 151. Y.Wang, Q.Lei, Z.Yan, C.Shen, N.Wang. Biochem. Pharmacol., 155, 510 2018); https://doi.org/10.1016/j.bcp.2018.07.035
- 152. Y.Wang, Q.Lei, C.Shen, N.Wang. *Cancer Chemother: Pharmacol.*, 88 (2), 289 (2021); https://doi.org/10.1007/s00280-021-04283-5
- N.Krishna Moorthy, O.Seifert, S.Eisler, S.Weirich, R.E.Kontermann, M.Rehm, G.Fullstone. *Molecules*, 26 (24), 7582 (2021); https://doi.org/10.3390/molecules26247582
- A.C.Di Polidoro, A.Cafarchio, D.Vecchione, P.Donato, F.De Nola, E.Torino. *Molecules*, **27** (19), 6696 (2022); https://doi.org/10.3390/molecules27196696
- Z.-L.Liu, H.-H.Chen, L.-L.Zheng, L.-P.Sun, L.Shi. Sig. Transduct. Target Ther., 8 (1), 198 (2023); https://doi.org/10.1038/s41392-023-01460-1
- 156. R.Huegel, P.Velasco, M.De La Luz Sierra, E.Christophers, J.M.Schröder, T.Schwarz, G.Tosato, B.Lange-Asschenfeldt. *J. Invest. Dermatol.*, **127** (1), 65 2007); https://doi.org/10.1038/sj.jid.5700484
- J.Fan, Z.Wang, L.Huang, Y.Shen. Protein Expression Purif., 125, 68 2016); https://doi.org/10.1016/j.pep.2015.09.007
- 158. P.Rozga, D.Kloska, S.Pawlak, M.Teska-Kaminska, M.Galazka, K.Bukato, A.Pieczykolan, A.Jaworski, A.Molga-Kaczmarska, A.Kopacz, B.Badyra, N.Kachamakova-Trojanowska, O.Zolnierkiewicz, M.Targosz-Korecka, K.Poleszak, M.Szymanik, B.Zerek, J.Pieczykolan, A.Jozkowicz, A.Grochot-Przeczek. *Int. J. Cancer*, **147** (4), 1117 (2020); https://doi.org/10.1002/ijc.32845
- K.Piechna, A.Żołyniak, E.Jabłońska, M.Noyszewska-Kania, M.Szydłowski, B.Żerek, M.Kulecka, I.Rumieńczyk, M.Mikula, P.Juszczyński. *Front. Oncol.*, **12**, 1048741 (2022); https://doi.org/10.3389/fonc.2022.1048741
- M.Kopczynski, M.Statkiewicz, M.Cybulska, U.Kuklinska, K.Unrug-Bielawska, Z.Sandowska-Markiewicz, A.Grochowska, M.Gajewska, M.Kulecka, J.Ostrowski, M.Mikula. *Int. J. Mol. Sci.*, **22** (6), 3160 (2021); https://doi.org/10.3390/ijms22063160
- 161. A.V.Yagolovich, A.A.Artykov, A.A.Isakova, Y.V.Vorontsova, D.A.Dolgikh, M.P.Kirpichnikov, M.E.Gasparian. *Int. J. Mol. Sci.*, 23 (11), 5860 (2022); https://doi.org/10.3390/ijms23115860
- 162. A.A.Isakova, A.A.Artykov, E.A.Plotnikova, G.V.Trunova, V.A.Khokhlova, A.A.Pankratov, M.L.Shuvalova, D.V.Mazur, N.V.Antipova, M.I.Shakhparonov, D.A.Dolgikh, M.P.Kirpichnikov, M.E.Gasparian, A.V.Yagolovich. *Int. J. Biol. Macromol.*, 255, 128096 (2024); https://doi.org/10.1016/j.ijbiomac.2023.128096
- 163. M.E.Gasparian, M.L.Bychkov, A.V.Yagolovich, D.A.Dolgikh, M.P.Kirpichnikov. *Biochem. Moscow*, **80** (8), 1080 (2015); https://doi.org/10.1134/S0006297915080143
- 164. P.Monnier, R.Vigouroux, N.Tassew. Antibodies, 2 (2), 193 (2013); https://doi.org/10.3390/antib2020193
- 165. P.Muñoz-López, R.M.Ribas-Aparicio, E.I.Becerra-Báez, K.Fraga-Pérez, L.F.Flores-Martínez, A.A.Mateos-Chávez, R.Luria-Pérez. *Cancers*, 14 (17), 4206 2022); https://doi.org/10.3390/cancers14174206
- R.V.Kholodenko, D.V.Kalinovsky, I.I.Doronin,
   E.D.Ponomarev, I.V.Kholodenko. *Curr. Med. Chem.*, 26 (3), 396 (2019); https://doi.org/10.2174/0929867324666170817152554

- M.De Bruyn, E.Bremer, W.Helfrich. *Cancer Lett.*, **332** (2), 175 (2013); https://doi.org/10.1016/j.canlet.2010.11.006
- H.Wajant, D.Moosmayer, T.Wüest, T.Bartke, E.Gerlach, U.Schönherr, N.Peters, P.Scheurich, K.Pfizenmaier. *Oncogene*, 20 (30), 4101 (2001); https://doi.org/10.1038/sj.onc.1204558
- E.Bremer, J.Kuijlen, D.Samplonius, H.Walczak, L.De Leij, W.Helfrich. *Int. J. Cancer*, **109** (2), 281 (2004); https://doi.org/10.1002/ijc.11702
- E.Bremer, D.Samplonius, B.-J.Kroesen, L.Van Genne, L.De Leij, W.Helfrich. *Neoplasia*, 6 (5), 636 (2004); https://doi.org/10.1593/neo.04229
- E.Bremer, D.F.Samplonius, M.Peipp, L.Van Genne, B.-J.Kroesen, G.H.Fey, M.Gramatzki, L.F.M.H.De Leij, W.Helfrich. *Cancer Res.*, 65 (8), 3380 (2005); https://doi.org/10.1158/0008-5472.CAN-04-2756
- 172. E.Levantini, G.Maroni, M.Del Re, D.G.Tenen. Seminars Cancer Biol., 85, 253 (2022); https://doi.org/10.1016/j.semcancer.2022.04.002
- 173. E.Bremer, D.F.Samplonius, L.Van Genne, M.H.Dijkstra, B.J.Kroesen, L.F.M.H.De Leij, W.Helfrich. J. Biol. Chem., 280 (11), 10025 (2005); https://doi.org/10.1074/jbc.M413673200
- 174. E.Bremer, G.M.Van Dam, M.De Bruyn, M.Van Riezen, M.Dijkstra, G.Kamps, W.Helfrich, H.Haisma. *Mol. Ther.*, **16** (12), 1919 (2008); https://doi.org/10.1038/mt.2008.203
- 175. O.Hammer. *mAbs*, **4** (5), 571 (2012); https://doi.org/10.4161/mabs.21338
- 176. J.Stieglmaier, E.Bremer, C.Kellner, T.M.Liebig, B.Ten Cate, M.Peipp, H.Schulze-Koops, M.Pfeiffer, H.-J.Bühring, J.Greil, F.Oduncu, B.Emmerich, G.H.Fey, W.Helfrich. *Cancer Immunol. Immunother.*, **57** (2), 233 (2008); https://doi.org/10.1007/s00262-007-0370-8
- B.Ten Cate, E.Bremer, M.De Bruyn, T.Bijma, D.Samplonius, M.Schwemmlein, G.Huls, G.Fey, W.Helfrich. *Leukemia*, 23 (8), 1389 (2009); https://doi.org/10.1038/leu.2009.34
- J.E.Maakaron, J.Rogosheske, M.Long, V.Bachanova, A.S.Mims. J. Clin. Pharmacol., 61 (1), 7 (2021); https://doi.org/10.1002/jcph.1730
- 179. M.De Bruyn, A.A.Rybczynska, Y.Wei, M.Schwenkert, G.H.Fey, R.A.Dierckx, A.Van Waarde, W.Helfrich, E.Bremer. *Mol. Cancer*, 9 (1), 301 (2010); https://doi.org/10.1186/1476-4598-9-301
- 180. K.M.Ilieva, A.Cheung, S.Mele, G.Chiaruttini, S.Crescioli, M.Griffin, M.Nakamura, J.F.Spicer, S.Tsoka, K.E.Lacy, A.N.J.Tutt, S.N.Karagiannis. *Front. Immunol.*, 8, 1911 (2018); https://doi.org/10.3389/fimmu.2017.01911
- 181. Y.He, D.Hendriks, R.Van Ginkel, D.Samplonius, E.Bremer, W.Helfrich. J. Invest. Dermatol., **136** (2), 541 (2016); https://doi.org/10.1016/j.jid.2015.11.009
- 182. M.De Bruyn, Y.Wei, V.R.Wiersma, D.F.Samplonius, H.G.Klip, A.G.J.Van Der Zee, B.Yang, W.Helfrich, E.Bremer. *Clin. Cancer Res.*, **17** (17), 5626 2011); https://doi.org/10.1158/1078-0432.CCR-11-0303
- 183. E.Luis, A.Anaya-Hernández, P.León-Sánchez, M.L.Durán-Pastén. *Int. J. Mol. Sci.*, 23 (15), 8458 (2022); https://doi.org/10.3390/ijms23158458
- F.Hartung, W.Stühmer, L.A.Pardo. *Mol. Cancer*, **10** (1), 109 (2011); https://doi.org/10.1186/1476-4598-10-109
- 185. F.Hartung, L.A.Pardo. *Eur. Biophys. J.*, **45** (7), 709 (2016); https://doi.org/10.1007/s00249-016-1149-7
- 186. X.Jia, B.Yan, X.Tian, Q.Liu, J.Jin, J.Shi, Y.Hou. Int. J. Biol. Sci., 17 (13), 3281 (2021); https://doi.org/10.7150/ijbs.60782
- 187. V.R.Wiersma, Y.He, D.F.Samplonius, R.J.Van Ginkel, J.Gerssen, P.Eggleton, J.Zhou, E.Bremer, W.Helfrich. *Br. J. Haematol.*, **164** (2), 304 (2014); https://doi.org/10.1111/bjh.12617
- 188. H.Ma, I.S.Padmanabhan, S.Parmar, Y.Gong. J. Hematol. Oncol., 12 (1), 41 (2019); http://linkinglobal.com/solid.com/soli
- https://doi.org/10.1186/s13045-019-0726-5
- V.R.Wiersma, M.De Bruyn, C.Shi, M.J.Gooden, M.C.Wouters, D.F.Samplonius, D.Hendriks, H.W.Nijman,

Y.Wei, J.Zhou, W.Helfrich, E.Bremer. *mAbs.*, **7** (2), 321 (2015); https://doi.org/10.1080/19420862.2015.1007811

- R.H.Vonderheide. Annu. Rev. Med., 71 (1), 47 (2020); https://doi.org/10.1146/annurev-med-062518-045435
- 191. M.El-Mesery, J.Trebing, V.Schäfer, D.Weisenberger, D.Siegmund, H.Wajant. *Cell Death Dis.*, 4 (11), e916 (2013); https://doi.org/10.1038/cddis.2013.402
- C.Dostert, M.Grusdat, E.Letellier, D.Brenner. *Physiol. Rev.*, **99** (1), 115 (2019); https://doi.org/10.1152/physrev.00045.2017
- J.Trebing, M.El-Mesery, V.Schäfer, D.Weisenberger, D.Siegmund, K.Silence, H.Wajant. *Cell Death Dis.*, 5 (1), e1035 (2014); https://doi.org/10.1038/cddis.2013.555
- 194. C.Yan, S.Li, Z.Li, H.Peng, X.Yuan, L.Jiang, Y.Zhang, D.Fan, X.Hu, M.Yang, D.Xiong. *Mol. Pharmaceutics*, **10** (1), 142 (2013); https://doi.org/10.1021/mp300261e
- 195. G.Pavlasova, M.Mraz. *Haematologica*, **105** (6), 1494 (2020); https://doi.org/10.3324/haematol.2019.243543
- 196. C.-T.Kuan, K.Wakiya, J.E.Herndon, E.S.Lipp, C.N.Pegram, G.J.Riggins, A.Rasheed, S.E.Szafranski, R.E.McLendon, C.J.Wikstrand, D.D.Bigner. *BMC Cancer*, **10** (1), 468 (2010); https://doi.org/10.1186/1471-2407-10-468
- 197. L.-H.Wang, C.-W.Ni, Y.-Z.Lin, L.Yin, C.-B.Jiang, C.-T.Lv, Y.Le, Y.Lang, C.-Y.Zhao, K.Yang, B.-H.Jiao, J.Yin. *Tumor Biol.*, **35** (2), 1157 (2014); https://doi.org/10.1007/s13277-013-1155-7
- 198. A.Rehman, Y.Huang, H.Wan. *Life*, **11** (7), 621 (2021); https://doi.org/10.3390/life11070621
- 199. M.Kouno, C.Lin, N.M.Schechter, D.Siegel, X.Yang, J.T.Seykora, J.R.Stanley. J. Invest. Dermatol., **133** (9), 2212 (2013); https://doi.org/10.1038/jid.2013.85
- 200. A.V.R.Kornepati, R.K.Vadlamudi, T.J.Curiel. Nat. Rev. Cancer, 22 (3), 174 (2022); https://doi.org/10.1038/s41568-021-00431-4
- 201. D.Hendriks, Y.He, I.Koopmans, V.R.Wiersma, R.J.van Ginkel, D.F.Samplonius, W.Helfrich, E.Bremer. *OncoImmunology*, 5 (8), e1202390 (2016); https://doi.org/10.1080/2162402X.2016.1202390
- Y.Zhu, N.Bassoff, C.Reinshagen, D.Bhere, M.O.Nowicki, S.E.Lawler, J.Roux, K.Shah. *Sci. Rep.*, 7 (1), 2602 (2017); https://doi.org/10.1038/s41598-017-2483-9
- 203. X.Zou, X.-Y.Tang, Z.-Y.Qu, Z.-W.Sun, C.-F.Ji, Y.-J.Li, S.-D.Guo. Int. J. Biol. Macromol., 202, 539 (2022); https://doi.org/10.1016/j.ijbiomac.2022.01.113
- 204. Z.Tao, H.Yang, Q.Shi, Q.Fan, L.Wan, X.Lu. *Theranostics*, 7 (8), 2261 (2017); ttps://doi.org/10.7150/thno.19091
- 205. R.Li, Z.Li, Y.Feng, H.Yang, Q.Shi, Z.Tao, J.Cheng, X.Lu. *Apoptosis*, **25** (1), 105 (2020); https://doi.org/10.1007/s10495-019-01583-3
- 206. Z.Tao, Y.Liu, H.Yang, Y.Feng, H.Li, Q.Shi, S.Li, J.Cheng, X.Lu. *Biomacromolecules*, **21** (10), 4017 (2020); https://doi.org/10.1021/acs.biomac.0c00785
- 207. Y.Zhang. *Pharmacol. Rev.*, **75** (6), 1218 (2023); https://doi.org/10.1124/pharmrev.123.000906
- B.Schneider, S.Münkel, A.Krippner-Heidenreich, I.Grunwald, W.S.Wels, H.Wajant, K.Pfizenmaier, J.Gerspach. *Cell Death Dis.*, 1 (8), e68 (2010); https://doi.org/10.1038/cddis.2010.45
- M.Siegemund, N.Pollak, O.Seifert, K.Wahl, K.Hanak,
   A.Vogel, A.K.Nussler, D.Göttsch, S.Münkel, H.Bantel,
   R.E.Kontermann, K.Pfizenmaier. *Cell Death Dis.*, 3 (4), e295 (2012); https://doi.org/10.1038/cddis.2012.29
- Y.Möller, M.Siegemund, S.Beyes, R.Herr, D.Lecis, D.Delia, R.Kontermann, T.Brummer, K.Pfizenmaier, M.A.Olayioye. *PLoS ONE*, 9 (9), e107165 (2014); https://doi.org/10.1371/journal.pone.0107165
- O.Seifert, A.Plappert, S.Fellermeier, M.Siegemund, K.Pfizenmaier, R.E.Kontermann. *Mol. Cancer Ther.*, **13** (1), 101 (2014); https://doi.org/10.1158/1535-7163.MCT-13-0396
- 212. M.Hutt, L.Marquardt, O.Seifert, M.Siegemund, I.Müller, D.Kulms, K.Pfizenmaier, R.E.Kontermann. *Mol. Cancer Ther.*, 16 (12), 2792 (2017); https://doi.org/10.1158/1535-7163.MCT-17-0551

- M.Hutt, S.Fellermeier-Kopf, O.Seifert, L.C.Schmitt, K.Pfizenmaier, R.E.Kontermann. *Oncotarget.*, 9 (13), 11322 (2018); https://doi.org/10.18632/oncotarget.24379
- 214. F.Morandi, A.L.Horenstein, F.Costa, N.Giuliani, V.Pistoia, F.Malavasi. *Front. Immunol.*, 9, 2722 (2018); https://doi.org/10.3389/fimmu.2018.02722
- R.De Luca, P.Kachel, K.Kropivsek, B.Snijder, M.G.Manz, D.Neri. Protein Eng. Design Selection, 31 (5), 173 (2018); https://doi.org/10.1093/protein/gzy015
- F.Hartung, T.Krüwel, X.Shi, K.Pfizenmaier, R.Kontermann, P.Chames, F.Alves, L.A.Pardo. *Front. Pharmacol.*, **11**, 686 (2020); https://doi.org/10.3389/fphar.2020.00686
- 217. D.Winterberg, L.Lenk, M.Oßwald, F.Vogiatzi, C.L.Gehlert, F.-S.Frielitz, K.Klausz, T.Rösner, T.Valerius, A.Trauzold, M.Peipp, C.Kellner, D.M.Schewe. J. Clin. Med., 10 (12), 2634 (2021); https://doi.org/10.3390/jcm10122634
- 218. M.Kundu, Y.E.Greer, J.L.Dine, S.Lipkowitz. *Cells*, **11** (23), 3717 (2022); https://doi.org/10.3390/cells11233717
- 219. P.J.Kaplan-Lefko, J.D.Graves, S.J.Zoog, Y.Pan, J.Wall, D.G.Branstetter, J.Moriguchi, A.Coxon, J.N.Huard, R.Xu, M.L.Peach, M.L.Peach, G.Juan, S.Kaufman, Q.Chen, A.Bianchi, J.J.Kordich, M.Ma, I.N.Foltz, B.C.Gliniak. *Cancer Biol. Ther.*, **9** (8), 618 (2010); https://doi.org/10.4161/cbt.9.8.11264
- N.S. Wilson, B.Yang, A.Yang, S.Loeser, S.Marsters, D.Lawrence, Y.Li, R.Pitti, K.Totpal, S.Yee, S.Ross, J.-M.Vernes, Y.Lu, C.Adams, R.Offringa, B.Kelley, S.Hymowitz, D.Daniel, G.Meng, A.Ashkenazi. *Cancer Cell*, **19** (1), 101 (2011); https://doi.org/10.1016/j.ccr.2010.11.012
- 221. F.Li, J.V.Ravetch. Proc. Natl. Acad. Sci. USA, 109 (27), 10966 (2012); https://doi.org/10.1073/pnas.1208698109
- M.B.Overdijk, K.Strumane, F.J.Beurskens, A.Ortiz Buijsse, C.Vermot-Desroches, B.S.Vuillermoz, T.Kroes, B.De Jong, N.Hoevenaars, R.G.Hibbert, A.Lingnau, U.Forssmann, J.Schuurman, P.W.H.I.Parren, R.N.De Jong, E.C.W.Breij. *Mol. Cancer Ther.*, **19** (10), 2126 (2020); https://doi.org/10.1158/1535-7163.MCT-20-0044
- 223. H.J.Van Der Horst, A.T.Gelderloos, M.E.D.Chamuleau, E.C.W.Breij, S.Zweegman, I.S.Nijhof, M.B.Overdijk, T.Mutis. *Blood Adv.*, 5 (8), 2165 2021); https://doi.org/10.1182/bloodadvances.2020003731
- N.J.Lakhani, D.Berz, V.Andrianov, W.Crago, M.Holcomb, A.Hussain, C.Veldstra, J.Kalabus, B.O'Neill, L.Senne, E.Rowell, A.B.Heidt, K.M.Willis, B.P.Eckelman. *Clin. Cancer Res.*, 29 (16), 2988 (2023); https://doi.org/10.1158/1078-0432.CCR-23-0974
- 225. J.S.Michaelson, S.J.Demarest, B.Miller, A.Amatucci,
  W.B.Snyder, X.Wu, F.Huang, S.Phan, S.Gao, A.Doern,
  G.K.Farrington, A.A.Lugovskoy, I.Joseph, V.Bailly, X.Wang,
  E.Garber, J.Browning, S.M.Glaser. *mAbs*, 1 (2), 128 2009);
  https://doi.org/10.4161/mabs.1.2.7631
- P.Brünker, K.Wartha, T.Friess, S.Grau-Richards, I.Waldhauer, C.F.Koller, B.Weiser, M.Majety, V.Runza, H.Niu, K.Packman, N.Feng, S.Daouti, R.J.Hosse, E.Mössner, T.G.Weber, F.Herting, W.Scheuer, H.Sade, C.Shao, B.Liu, P.Wang, G.Xu, S.Vega-Harring, C.Klein, K.Bosslet, P.Umaña. *Mol. Cancer Ther.*, **15** (5), 946 (2016); https://doi.org/10.1158/1535-7163.MCT-15-0647
- A.Satta, D.Mezzanzanica, F.Caroli, B.Frigerio, M.D.Nicola, R.E.Kontermann, F.Iacovelli, A.Desideri, A.Anichini, S.Canevari, A.M.Gianni, M.Figini. *mAbs*, 19420862.2018.1494105 (2018); https://doi.org/10.1080/19420862.2018.1494105
- G.Shivange, K.Urbanek, P.Przanowski, J.S.A.Perry, J.Jones, R.Haggart, C.Kostka, T.Patki, E.Stelow, Y.Petrova, D.Llaneza, M.Mayo, K.S.Ravichandran, C.N.Landen, S.Bhatnagar, J.Tushir-Singh. *Cancer Cell.*, 34 (2), 331 (2018); https://doi.org/10.1016/j.ccell.2018.07.005
- 229. K.P.Papadopoulos, R.Isaacs, S.Bilic, K.Kentsch, H.A.Huet, M.Hofmann, D.Rasco, N.Kundamal, Z.Tang, J.Cooksey,

A.Mahipal. *Cancer Chemother. Pharmacol.*, **75** (5), 887 (2015); https://doi.org/10.1007/s00280-015-2712-0

- J.M.García-Martínez, S.Wang, C.Weishaeupl, A.Wernitznig, P.Chetta, C.Pinto, J.Ho, D.Dutcher, P.N.Gorman, R.Kroe-Barrett, J.Rinnenthal, C.Giragossian, M.A.Impagnatiello, I.Tirapu, F.Hilberg, N.Kraut, M.Pearson, K.P.Kuenkele. *Mol. Cancer Ther.*, 20 (1), 96 (2021); https://doi.org/10.1158/1535-7163.MCT-20-0253
- 231. F.Jacobsen, R.Pushpadevan, F.Viehweger, M.Freytag, R.Schlichter, N.Gorbokon, F.Büscheck, A.M.Luebke, D.Putri, M.Kluth, C.Hube-Magg, A.Hinsch, D.Höflmayer, C.Fraune, C.Bernreuther, P.Lebok, G.Sauter, S.Minner, S.Steurer, R.Simon, E.Burandt, D.Dum, F.Lutz, A.H.Marx, T.Krech, T.S.Clauditz. *Path. Res. Practice*, **256**, 155175 (2024); https://doi.org/10.1016/j.prp.2024.155175
- 232. S.Dhillon. *Drugs*, **84** (4), 459 (2024); https://doi.org/10.1007/s40265-024-02004-9
- 233. Y.Guo, A.Roohullah, J.Xue, W.Zhao, M.Aghmesheh, D.Martin, Y.Zhou, C.Gao, Y.Yang, D.-Z.Xu, J.Li. *Cancer Res.*, **82** (12\_Supplement), 6180 (2022); https://doi.org/10.1158/1538-7445.AM2022-6180
- 234. P.LoRusso, M.J.Ratain, T.Doi, D.W.Rasco, M.J.A.De Jonge, V.Moreno, B.A.Carneiro, L.A.Devriese, A.Petrich, D.Modi, S.Morgan-Lappe, S.Nuthalapati, M.Motwani, M.Dunbar, J.Glasgow, B.C.Medeiros, E.Calvo. *Invest. New Drugs*, **40** (4), 762 (2022); https://doi.org/10.1007/s10637-022-01247-1
- J.Bendell, J.-Y.Blay, P.Cassier, T.Bauer, C.Terret, C.Mueller, A.Morel, E.Chesne, Z.Xu, J.Tessier, M.Ceppi, I.James, S.Wilson, E.Quackenbush, M.Ochoa De Olza, J.Tabernero, M.De Miguel, E.Calvo. *Mol. Cancer Ther.*, **17** (1\_Suppl.), A092 (2018);
- https://doi.org/10.1158/1535-7163.TARG-17-A092
  236. J.J.Harding, R.-D.Hofheinz, E.Elez, Y.Kuboki, D.W.Rasco, M.Cecchini, L.Shen, M.He, S.Archuadze, N.Chhaya, S.Pant. *J. Clin. Oncol.*, 41 (4\_suppl), 115 (2023);
  https://doi.org/10.1200/JCO.2023.41.4 suppl.115